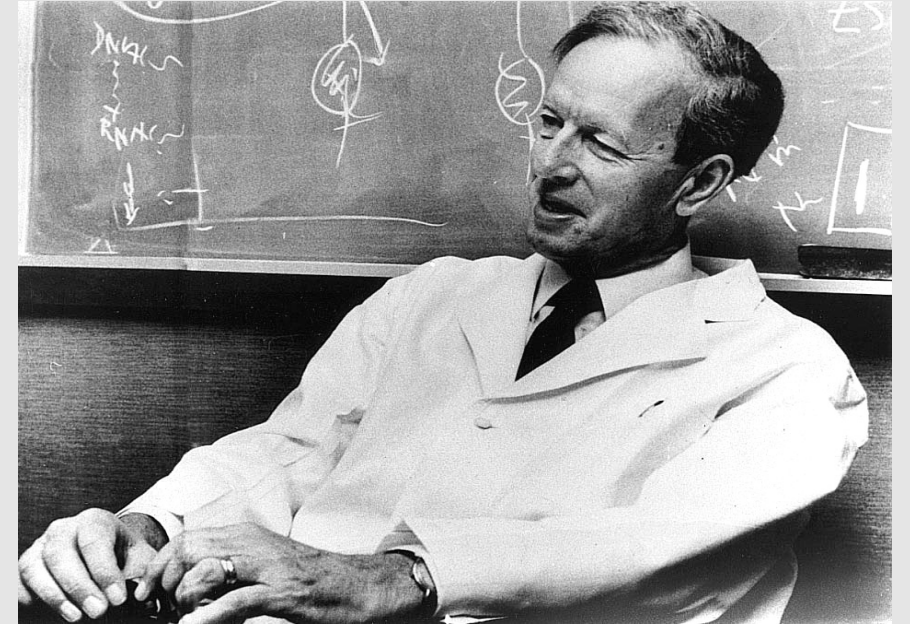


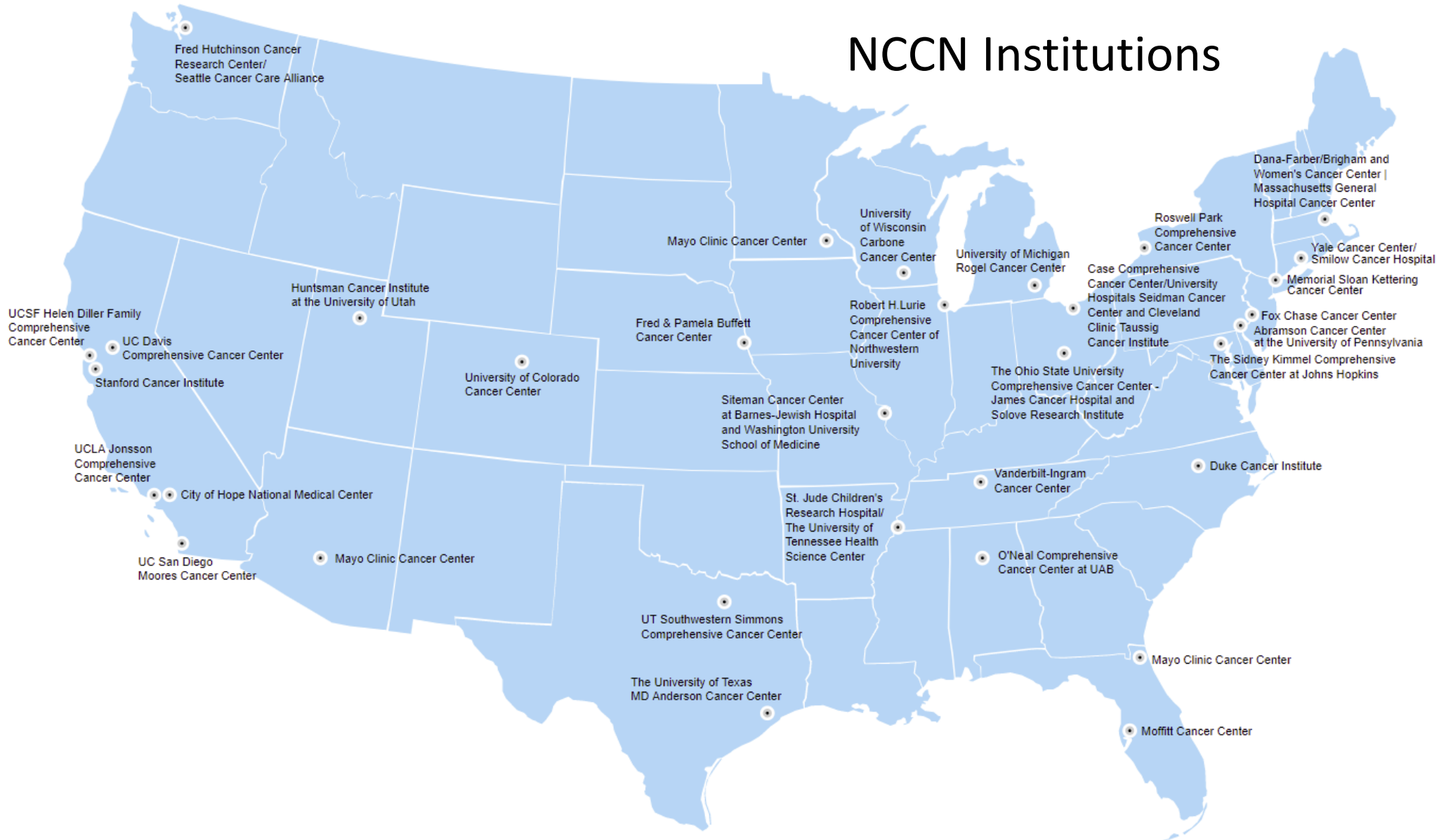
NCCN Guidelines for LPL/Waldenstrom's Macroglobulinemia, IgM MGUS and IgM Related Disorders.

Steven P. Treon MD, PhD, FACP, FRCP
Professor of Medicine
Harvard Medical School
Director
Bing Center for Waldenstrom's Macroglobulinemia
Dana Farber Cancer Institute



This presentation was made at the annual IWMF virtual meeting and is intended for educational purposes only and is based on best available information in the opinion of the presenter at the time of the presentation. The information presented is not for specific patient advise. Patients should consult their physician for specific information relative to their ongoing management and treatment.

NCCN Institutions





WHO CRITERIA FOR LYMPHOPLASMACYTIC LYMPHOMA AND WALDENSTRÖM MACROGLOBULINEMIA

- **Lymphoplasmacytic lymphoma:**
 - ▶ Neoplasm of small B lymphocytes, plasmacytoid lymphocytes, and plasma cells
 - ▶ Usually involving bone marrow and sometimes lymph nodes and spleen
 - ▶ Does not fulfill criteria of any other small B-cell lymphoid neoplasm that may also have plasmacytic differentiation

Reproduced with permission from Swerdlow SH, Campo E, Harris NL, et al. World Health Organization Classification of Tumours of Haematopoietic and Lymphoid Tissues, revised 4th edition. IARC, Lyon 2017.

- **Waldenström macroglobulinemia:**
 - ▶ Lymphoplasmacytic lymphoma with bone marrow involvement and IgM monoclonal gammopathy of any concentration

Adapted with permission. Owen RG, Treon SP, Al-Katib A, et al. Clinicopathological Definition of Waldenström's Macroglobulinemia: Consensus Panel Recommendations from the Second International Workshop on Waldenström's Macroglobulinemia. *Semin Oncol.* 2003;30:110-115.

WALDENSTRÖM MACROGLOBULINEMIA INTERNATIONAL WORKSHOP CRITERIA

Proposed Criteria for the Diagnosis of Waldenström Macroglobulinemia

- IgM monoclonal gammopathy of any concentration
- Bone marrow infiltration by small lymphocytes, plasmacytoid cells, and plasma cells
- Diffuse, interstitial, or nodular pattern of bone marrow infiltration
- CD19+, CD20+, sIgM+; CD5, CD10, CD23 can be expressed in some cases of Waldenström macroglobulinemia and does not exclude diagnosis.

Reprinted with permission from Elsevier. Owen RG. Developing diagnostic criteria in Waldenström's macroglobulinemia. *Semin Oncol* 2003;30:196-200.

REVISED IPSS WALDENSTRÖM MACROGLOBULINEMIA SCORING SYSTEM

Criteria for the Diagnosis of Waldenström Macroglobulinemia (only at the time of initial treatment prognostication)

Table 1

	Points
Age <65	0
Age 66–75	1
Age >75	2
B2 microglobulin >4 mg/L	1
LDH >250 IU/L	1
Serum albumin <3.5 g/dL	1

Table 2

Score*	Stage
0	Very Low
1	Low
2	Intermediate
3	High
4–5	Very High

*Sum of total points in table 1

Adapted with permission from: Kastritis E, Morel P, Duhamel A, et al. A revised international prognostic score system for Waldenström's macroglobulinemia. *Leukemia* 2019;33:2654-2661.

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

NCCN Guidelines Version 1.2022

Waldenström Macroglobulinemia/ Lymphoplasmacytic Lymphoma

DIAGNOSIS

WORKUP^a

INDICATIONS FOR TREATMENT

- Essential^{b,c}**
- Hematopathology review of all slides with at least one paraffin block representative of the tumor (Rebiopsy if consult material is nondiagnostic)
 - Adequate tissue biopsy for immunophenotyping to establish diagnosis
 - ▶ Typical immunophenotype: CD19+, CD20+, sIgM+; CD5, CD10, CD23 may be positive in 10%–20% of cases and does not exclude diagnosis

Essential

- History and physical exam
- CBC, differential, platelet count
- Liver function tests (LFTs) as clinically indicated
- Peripheral blood smear
- Serum BUN/creatinine, electrolytes, albumin, calcium, serum uric acid, serum LDH, and beta-2 microglobulin
- Creatinine clearance (calculated or measured directly)
- Serum quantitative immunoglobulins, serum protein electrophoresis (SPEP), serum immunofixation electrophoresis (SIFE)
- Unilateral bone marrow aspirate and biopsy, including immunohistochemistry (IHC) and/or multi-parameter flow cytometry
- Chest/abdominal/pelvic CT with contrast when possible
- MYD88,^d L265P AS-PCR testing of bone marrow

Useful in Certain Circumstances

- Serum viscosity
- *CXCR4* gene mutation testing for patients being considered for ibrutinib^e
- Testing for hepatitis B (if rituximab planned), hepatitis C,^f and HIV
- Cryocrit^{f,g}
- Consider coagulation and/or von Willebrand disease testing if symptoms present (excess bruising or bleeding) or if clinically indicated
- Cold agglutinins
- Neurology consult^h
- Anti-MAG antibodies/anti-GM1^h
- Nerve conduction study (NCS)/electromyogram (EMG)^h
- Fat pad sampling and/or congo red staining of bone marrow for amyloid^h
- Retinal exam (if IgM ≥3.0 g/dL or if hyperviscosity is suspected)
- 24-h urine for total protein, urine protein electrophoresis (UPEP), and urine immunofixation electrophoresis (UIFE)
- Amyloid tissue subtyping with mass spectrometry, if indicated
- Brain/spine MRI, if CNS symptoms

Symptomsⁱ related to:

- Hyperviscosity
- Neuropathy
- Organomegaly
- Amyloidosis
- Cold agglutinin disease
- Cryoglobulinemia
- Anemia and other cytopenias associated with disease
- Bulky adenopathy
- B symptoms

[See Primary Treatment \(WM/LPL-2\)](#)



Peter Bing MD

ORIGINAL ARTICLE









MYD88 L265P Somatic Mutation in Waldenström's Macroglobulinemia

Steven P. Treon, M.D., Ph.D., Lian Xu, M.S., Guang Yang, Ph.D.,
Yangsheng Zhou, M.D., Ph.D., Xia Liu, M.D., Yang Cao, M.D.,
Patricia Sheehy, N.P., Robert J. Manning, B.S., Christopher J. Patterson, M.A.,
Christina Tripsas, M.A., Luca Arcaini, M.D., Geraldine S. Pinkus, M.D.,
Scott J. Rodig, M.D., Ph.D., Aliyah R. Sohani, M.D., Nancy Lee Harris, M.D.,
Jason M. Laramie, Ph.D., Donald A. Skifter, Ph.D., Stephen E. Lincoln, Ph.D.,
and Zachary R. Hunter, M.A.

**91% of WM patients positive by
Whole Genome Sequencing**

Treon et al, New Engl J Med 2012

MYD88 L265P in WM/IGM MGUS

		METHOD	TISSUE	WM	IGM MGUS
Treon		WGS/Sanger	BM CD19 ⁺	91%	10%
Xu		AS-PCR	BM CD19 ⁺	93%	54%
Gachard		PCR	BM	70%	
Varettoni		AS-PCR	BM	100%	47%
Landgren		Sanger	BM		54%
Jiminez		AS-PCR	BM	86%	87%
Poulain		PCR	BM CD19 ⁺	80%	
Argentou		PCR-RFLP	BM	92%	1/1 MGUS
Willenbacher		Sanger	BM	86%	
Mori		AS-PCR/BSiE1	BM	80%	
Ondrejka		AS-PCR	BM	100%	
Ansell		WES/AS-PCR	BM	97%	
Patkar		AS-PCR	BM	85%	

MYD88 Mutation Testing in B-cell LPDs

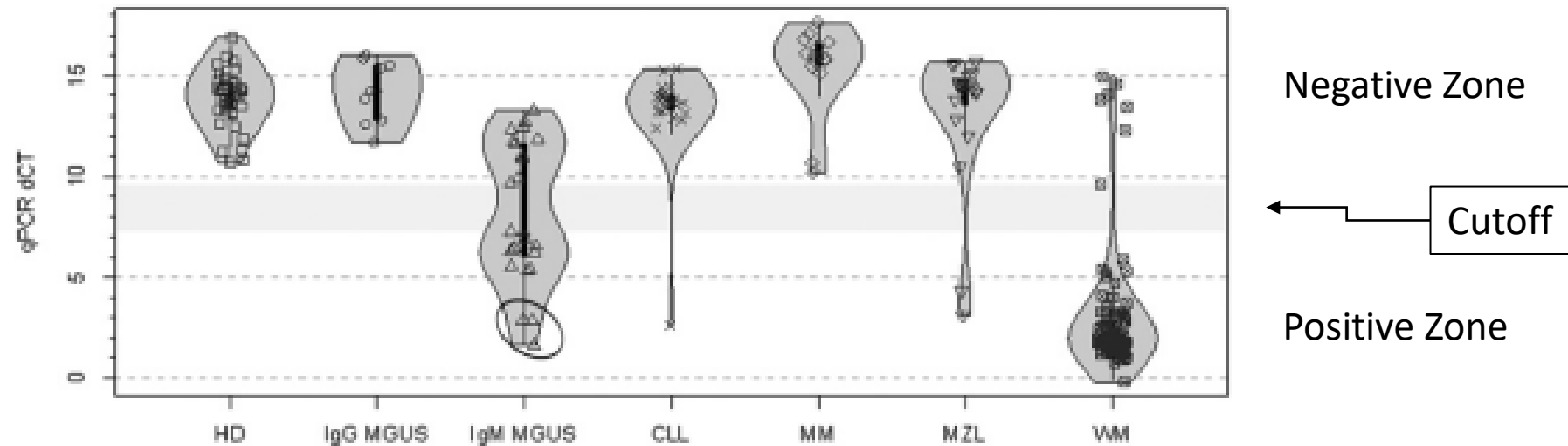
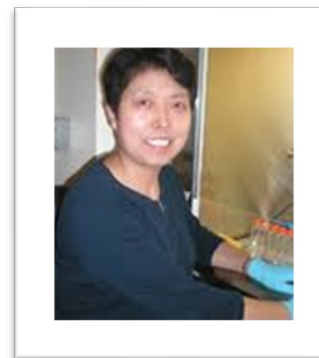


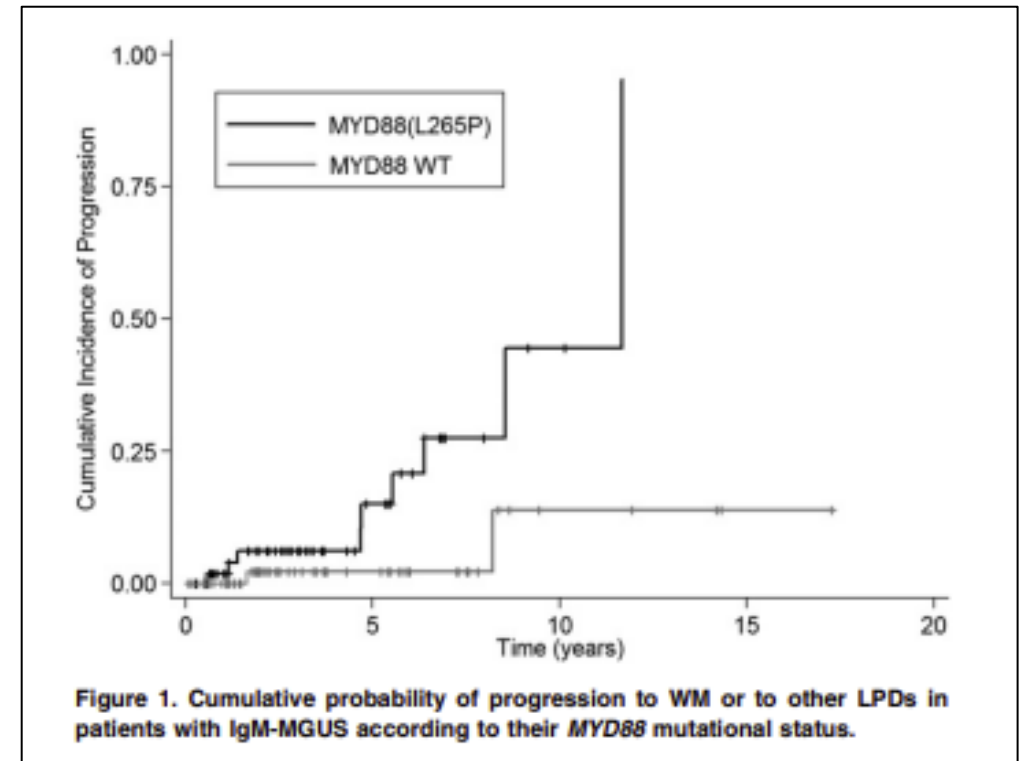
Figure 4. Real-time AS-PCR results for MYD88 L265P in samples from patients with WM, IgM MGUS, and other B-cell lymphoproliferative disorders. Violin plot representing AS-PCR differences in cycle threshold (ΔC_T). The span of grey area for each cohort represents the kernel density estimation of the sample distribution, and highlights the bimodal nature of the data. Box plots with interquartile ranges are shown in black with an overlay of the individual data points. Samples evaluated were from healthy donors (HD, $n = 40$); along with patients with IgG ($n = 9$) and IgM ($n = 24$) MGUS; CLL ($n = 26$); MM including 3 patients with IgM myeloma ($n = 14$); MZL ($n = 20$), and WM ($n = 104$). The light grey bar represents the distance between the highest positive (7.3), and lowest negative (9.6) sample ΔC_T values. Circled area depicts results for 3 IgM MGUS patients who progressed to WM.



Xu et al, Blood 2013

Progression of disease in IgM MGUS is related

- Absence of MYD88 and CXCR4 mutations in IGG and IGA MGUS.
- Mutated MYD88 and CXCR4 detected in 50 to 90% and 20% of IGM MGUS patients, respectively by AS-PCR.
- Presence of mutated MYD88 is an independent risk factor for progression.



Varettoni study: IGM MGUS patients subjected to AS-PCR. 71/138 Positive for MYD88 L265P. 11 progressed during follow-up (median 34 mos). 9/11 (82%) to WM, 8 of whom were MYD88 mutated; 2 to MZL (18%), 1 of whom was MYD88 mutated.

MYD88 status for classification of B-cell LPDS.

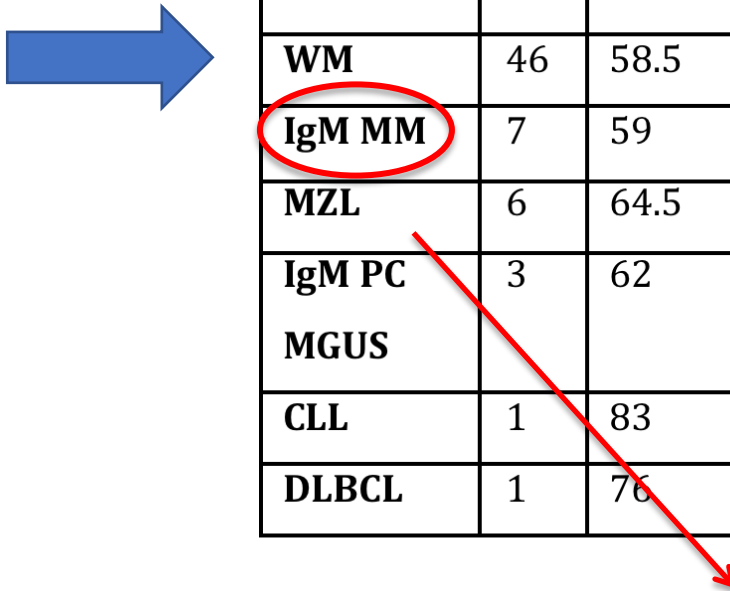
MYD88 status used to re-evaluate pathological diagnosis in 138 patients with B-cell LPDs. Mutation status was integrated with histologic and clinical data. Reclassification using molecular status is shown for five patients.

TABLE 2. Summary of Features of Reclassified Cases

Case No	Initial Diagnosis	Light-chain Restriction by Plasma Cells	Revised Diagnosis	Diagnostic Sample	Splenomegaly	Bone Marrow Involvement	Serum IgM Paraprotein (g/L)
1	NMZL	λ	LPL	LN	Absent	Present	24.0
2	BCL-NOS vs LPL	κ	LPL	BM	Absent	Present	11.0
3	BCL-NOS	κ	LPL	BM	Absent	Present	10.3
4	SMZL	κ	LPL	SPLEEN	Present	Present	7.3
5	SMZL	λ	LPL	SPLEEN	Present	Present	5.1

BM indicates bone marrow; LN, lymph node.

Differential Diagnosis of suspected non-MYD88 mutated LPL/WM



Diagnosis	N=	Age (yrs)	Gender (% male)	BM (%)	sIgM (mg/dL)	Hb (g/dL)	Adenopathy (%)	Splenomegaly (%)
WM	46	58.5	48	35	2,980	11.0	35	28
IgM MM	7	59	71	60	8,375	9.0	14	14
MZL	6	64.5	0	10	1,642	11.3	67	33
IgM PC MGUS	3	62	33	5	1,846	13.9	0	0
CLL	1	83	0	5	1,822	13.2	0	0
DLBCL	1	76	0	5	355	9.5	0	100

t(11;14); Cyclin D1 over-expression

N=64

MYD88 Testing for LPL/WM Extramedullary Pathology

Bing Neel Syndrome

Table I. Clinical, imaging and biological features of patients with BNS at entry into the study.

	Patient 1	Patient 2	Patient 3
Age (years)	36	59	68
Symptoms	Visual loss, drop attacks Absence of tumoural syndrome	Progressive leg palsy and blurry vision Absence of tumoural syndrome	Confusion, cognitive decline, dysphagia, Facial hemiparesis, hearing loss, difficulty walking, Absence of tumoural syndrome
Magnetic resonance imaging	Dura-mater, leptomeningeal and occipital involvement	Abnormal medullary and optic nerve signal	Predominantly occipital involvement
M-spike on SPE (g/l)	3-6	10-5	4
Blood lymphocytes counts ($\times 10^9/l$) (CD19% by flow cytometry)	2-6 (Not available)	3-6 (65)	1-4 (<1)
Protein concentration (g/l)*	2-83	0-66	5
Protein concentration index/M spike	0-78	0-06	1-25
Cellularity of CSF (/ul)	54	9	15
% of tumoural cells in CSF	51	80	75
Cytomorphology	Lymphoplasmacytic infiltration	Lymphoplasmacytic infiltration	Lymphoplasmacytic infiltration
Flow cytometry data on CSF	CD5-, CD10-, CD19 +, CD20 +, CD22 +, FMC7-, CD23-, CD38 +, IgM+, Kappa restriction	CD5-, CD10-, CD19 +, CD20 +, Kappa restriction	CD5 +, CD10-, CD19 +, CD20 +, Kappa restriction
Bone marrow involvement (%)	10	50	20
MYD88 L265P mutation			
Blood	Absent	Present	Absent
Bone marrow	Present	Present	Present
CSF	Present	Present	Present
Brain biopsy	Present	Not available	Not available

SPE serum protein electrophoresis; CSF: cerebrospinal fluid.

*normal range: 0-1-0-6 g/l.

Poulain et al, BJH 2014

Malignant Pleural Effusions

Table I. Clinical presentation and diagnostic test results of patients with malignant pleural effusions.

Patient	Age (years)	Gender	Symptoms	Imaging	Presentation	Serum IgM (g/l)	Pleural Fluid			MYD88 mutation		CXCR4 mutation	
							Cytology	Flow cytometry	IGH rearrangement	Bone marrow	Pleural fluid	Bone marrow	Pleural fluid
1	72	Female	SOB, cough	Bilateral moderate to large PE on right side	Progressive event off-therapy	19-6	No malignant cells	CD5+, CD10-, CD23+, CD19+, CD20+, polytypic LC	Polyclonal pattern	Present	Present	Present (S358X)	Not available
2	54	Female	SOB	Unilateral PE on right side	Progressive event while on ibrutinib	5-5	No malignant cells	CD5+, CD10-, CD23+, CD19+, CD20+, polytypic LC	Clonal pattern	Present	Present	Absent	Absent
3	73	Male	SOB, fatigue	Unilateral PE on left side	Progressive event while on ibrutinib	45-8	No malignant cells	CD5+, CD10-, CD23+, CD19+, CD20+, polytypic LC	Clonal pattern	Present	Present	Absent	Not available
4	68	Male	SOB, fatigue	Bilateral PE, left > right	Progressive event off-therapy	22-1	No malignant cells	CD5+, CD10-, CD23+, CD19+, CD20+, polytypic LC	Clonal pattern	Present	Present	Absent	Not available
5	70	Male	SOB, weight loss	Unilateral PE on left side	Progressive event off-therapy	24-7	No malignant cells	CD5+, CD10-, CD23+, CD19+, CD20+, polytypic LC	Clonal pattern	Present	Present	Absent	Absent
6	51	Male	SOB, fatigue	Unilateral PE on left side	Progressive event off-therapy	44-2	No malignant cells	CD5+, CD10-, CD23+, CD19+, CD20+, polytypic LC	Clonal pattern	Present	Present	Absent	Not available
7	75	Male	SOB	Unilateral PE on left side	Progressive event off-therapy	25-5	Malignant cells identified	CD5+, CD10-, CD23+, CD19+, CD20+, monotypic λ LC	Clonal pattern	Present	Present	Absent	Absent
8	69	Male	Cough on deep inspiration	Unilateral PE on left side	Progressive event off-therapy	22-2	No malignant cells	CD5+, CD10-, CD23+, CD19+, CD20+, polytypic LC	Not performed	Present	Present	Absent	Not available
9	69	Male	SOB, fatigue	Bilateral PE	Progressive event while on ibrutinib	6-1	Malignant cells identified	CD5+, CD10-, CD23+, CD19+, CD20+, monotypic λ LC	Not performed	Present*	Present*	Absent	Not available

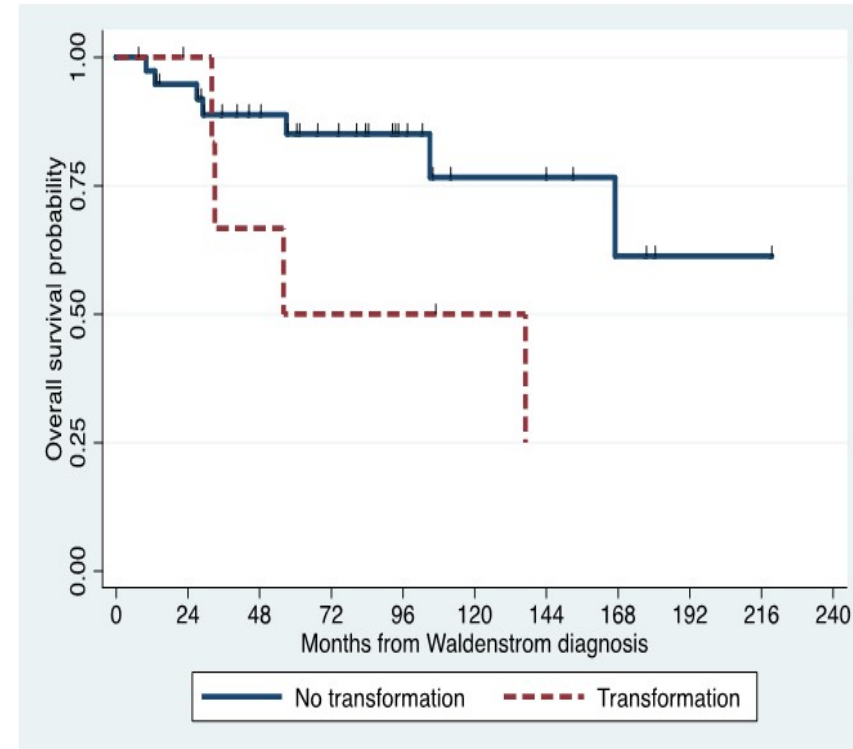
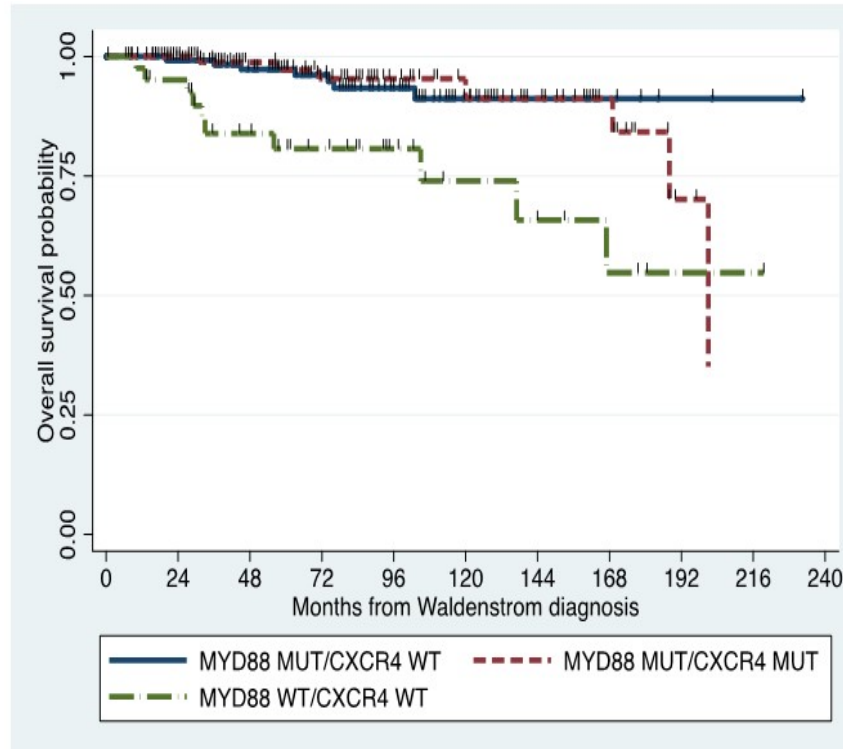
SOB, shortness of breath; PE, pleural effusion; LC, light chain.

*Patient 9 had MYD88 S243N detected; all other patients had MYD88 L265P identified.

Gustine et al, BJH 2016

Other studies: Himecke-Jiwa et al, Hematol Oncol 2018; Pan ST, et al, Pathol. Intl. 2019;

High risk of transformation and poorer survival accompany MYD88^{Wild-Type} LPL/WM

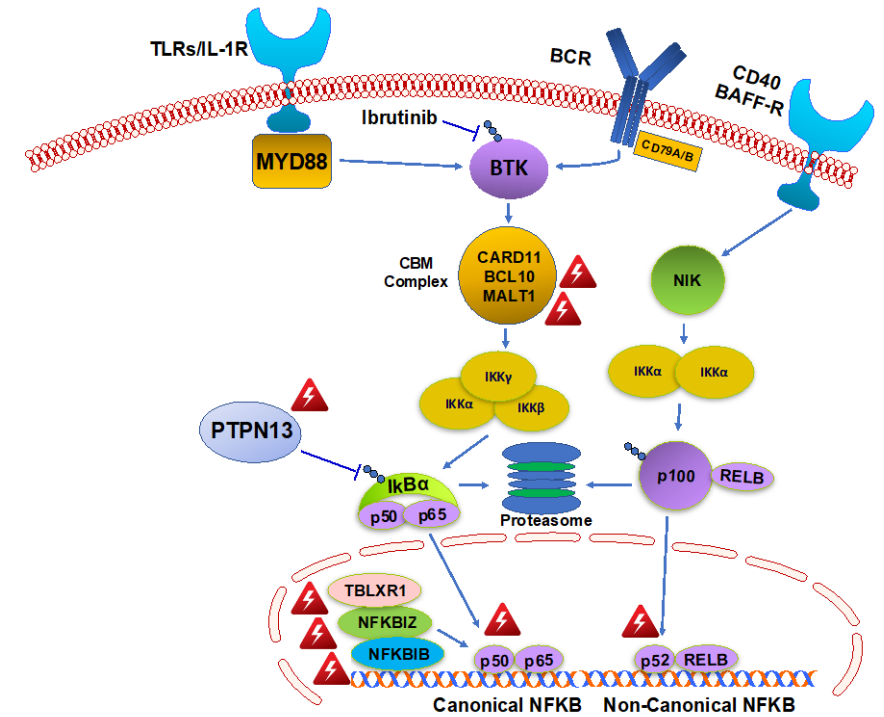
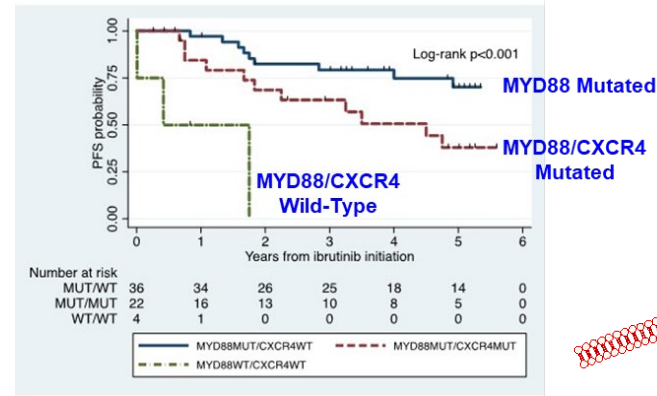


Transformation risk for MYD88 WT
(Odds ratio 23.3; 95% CI 4.2-233.8; $p < 0.001$).

Response to BTK-Inhibitors is lower in non-MYD88 mutated (MYD88 wild-type) WM.

Ibrutinib

	All Patients	MYD88 ^{MUT} CXCR4 ^{WT}	MYD88 ^{MUT} CXCR4 ^{MUT}	MYD88 ^{WT} CXCR4 ^{WT}	P-value
N=	63	36	22	4	N/A
Overall Response Rate-no. (%)	90.5%	100%	86.4%	50%	<0.01
Major Response Rate-no. (%)	79.4%	97.2%	68.2%	0%	<0.0001
Categorical responses					
Minor responses-no. (%)	11.1%	2.8%	18.2%	50%	<0.01
Partial responses-no. (%)	49.2%	50%	59.1%	0%	0.03
Very good partial responses-no. (%)	30.2%	47.2%	9.1%	0%	<0.01
Median time to response (months)					
Minor response (≥Minor response)	0.9	0.9	0.9	0.9	0.38
Major response (≥Partial response)	1.8	1.8	4.7	N/A	0.02

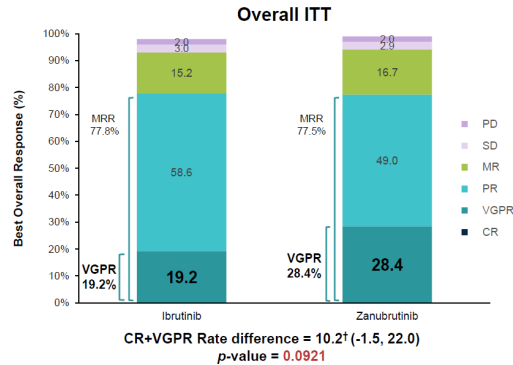


Zanubrutinib

MYD88 Mutated

ASPEN: Efficacy – Response by IRC (Data cutoff: 31 August 2019)

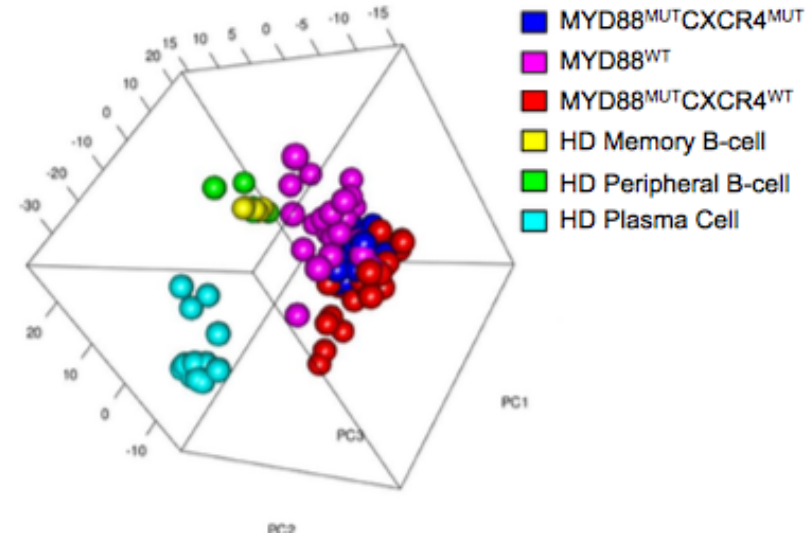
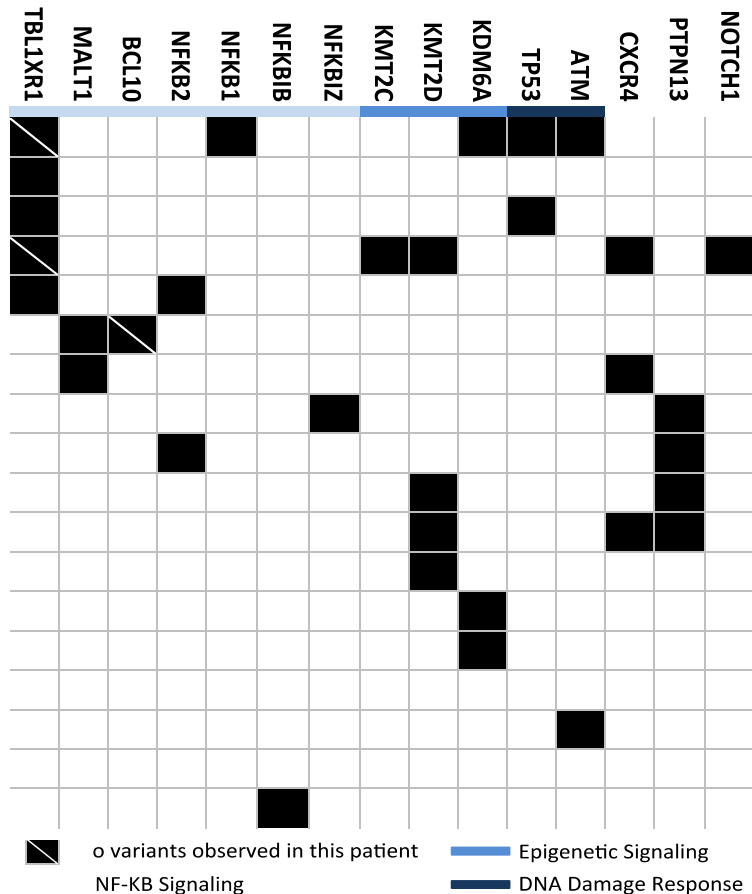
- Superiority in CR+VGPR rate compared to ibrutinib in relapsed/refractory population (primary study hypothesis) was not significant*



MYD88 Wild-Type

	N=28	N=	%
ORR		23	81%
Major (PR or better)		13	50%
VGPR		7	27%

Driver Mutations in WM patients without MYD88 mutations but gene expression overlaps with those who have the MYD88 mutation



Principal component analysis of top 500 high variance genes.

Hunter et al, Blood Adv. 2018

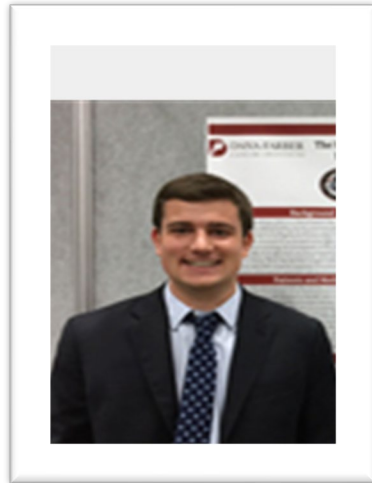
Challenges of MYD88 detection in WM: Comparison of AS-PCR vs. Next Generation Sequencing



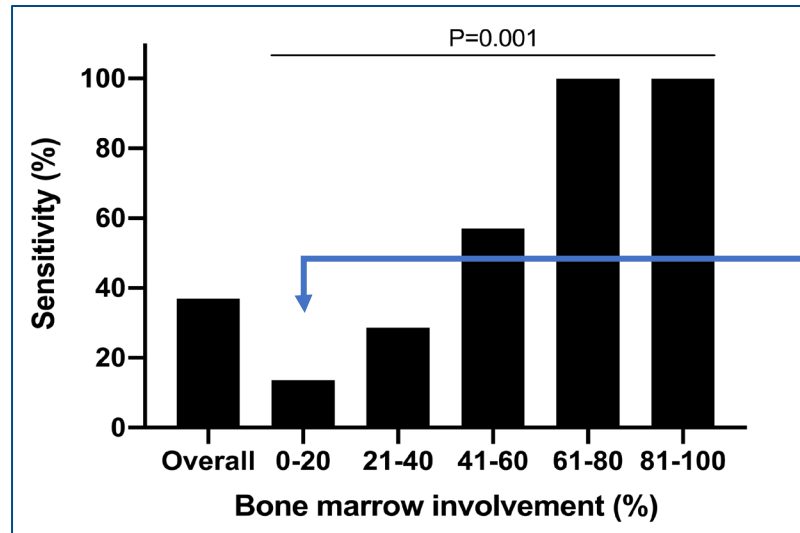
A. Kofides

	MYD88 L265P	
	AS-PCR	NGS
True Positive – no.	391	259
True Negative – no.	23	23
False Positive – no.	0	0
False Negative – no.	0	132
Concordance (κ) – %	Ref.	68 (0.19)
Sensitivity (95% CI) – %	Ref.	66 (61-71)
Specificity (95% CI) – %	Ref.	100 (83-100)
PPV (95% CI) – %	Ref.	100 (98-100)
NPV (95% CI) – %	Ref.	15 (10-22)

1 in 3 WM patients truly positive for MYD88 can be missed by NGS.



J. Gustine



WM patients with lower BM disease (<20%) involvement are more likely to be misclassified by NGS.

NCCN Guidelines Version 1.2022

Waldenström Macroglobulinemia/ Lymphoplasmacytic Lymphoma

DIAGNOSIS

WORKUP^a

INDICATIONS FOR TREATMENT

- Essential^{b,c}**
- Hematopathology review of all slides with at least one paraffin block representative of the tumor (Rebiopsy if consult material is nondiagnostic)
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 - ▶ Typical immunophenotype: CD19+, CD20+, sIgM+; CD5, CD10, CD23 may be positive in 10%–20% of cases and does not exclude diagnosis

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- History and physical exam
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- Consider coagulation and/or von Willebrand disease testing if symptoms present (excess bruising or bleeding) or if clinically indicated
- Cold agglutinins
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Symptomsⁱ related to:

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- Amyloidosis
- Cold agglutinin disease
- Cryoglobulinemia
- Anemia and other cytopenias associated with disease
- Bulky adenopathy
- B symptoms

[See Primary Treatment \(WM/LPL-2\)](#)

Discovery of CXCR4 mutations in WM -2013-

From www.bloodjournal.org by guest on May 8, 2018. For personal use only.

Plenary Paper

LYMPHOID NEOPLASIA

The genomic landscape of Waldenström macroglobulinemia is characterized by highly recurring MYD88 and WHIM-like CXCR4 mutations, and small somatic deletions associated with B-cell lymphomagenesis

Zachary R. Hunter,^{1,2} Lian Xu,¹ Guang Yang,¹ Yangsheng Zhou,¹ Xia Liu,¹ Yang Cao,¹ Robert J. Manning,¹ Christina Tripsas,¹ Christopher J. Patterson,¹ Patricia Sheehy,¹ and Steven P. Treon^{1,3}

¹Bing Center for Waldenström's Macroglobulinemia, Dana-Farber Cancer Institute, Boston, MA; ²Department of Pathology and Laboratory Medicine, Boston University School of Graduate Medical Sciences, Boston, MA; and ³Harvard Medical School, Boston, MA

Key Points

- Highly recurring mutations are present in WM, including MYD88 L265P, warts, hypogammaglobulinemia, infection, and myelokathexis-syndrome-like mutations in CXCR4, and ARID1A.
- Small, previously undetected CNAs affecting B-cell regulatory genes are highly prevalent in WM.

The genetic basis for Waldenström macroglobulinemia (WM) remains to be clarified. Although 6q losses are commonly present, recurring gene losses in this region remain to be defined. We therefore performed whole genome sequencing (WGS) in 30 WM patients, which included germline tumor sequencing for 10 patients. Validated somatic mutations occurring in >10% of patients included *MYD88*, *CXCR4*, and *ARID1A* that were present in 90%, 27%, and 17% of patients, respectively, and included the activating mutation L265P in MYD88 and warts, hypogammaglobulinemia, infection, and myelokathexis-syndrome-like mutations in CXCR4 that previously have only been described in the germline. WGS also delineated copy number alterations (CNAs) and structural variants in the 10 paired patients. The CXCR4 and CNA findings were validated in independent expansion cohorts of 147 and 30 WM patients, respectively. Validated gene losses due to CNAs involved *PRDM2* (93%), *BTG1* (87%), *HIVEP2* (77%), *MKLN1* (77%), *PLEKHG1* (70%), *LYN* (60%), *ARID1B* (50%), and *FOXP1* (37%). Losses in *PLEKHG1*, *HIVEP2*, *ARID1B*, and *BCLAF1* constituted the most common deletions within chromosome 6. Although no recurrent translocations were observed, in 2 patients deletions in 6q corresponded with translocation events. These studies evidence highly recurring somatic events, and provide a genomic basis for understanding the pathogenesis of WM. (*Blood*. 2014;123(11):1637-1646)



Hunter et al,
 Blood 2013

30-40% of WM patients carry CXCR4 mutations

MYD88 and CXCR4 Mutations

Clinical Presentation

Clinical Characteristics	MYD88 ^{L265P} CXCR4 ^{WT}	MYD88 ^{L265P} CXCR4 ^{WHIM/FS}	MYD88 ^{L265P} CXCR4 ^{WHIM/NS}	MYD88 ^{WT} CXCR4 ^{WT}
IgM	↑↑	↑↑	↑↑↑↑	↑
BM infiltration	↑↑↑	↑↑	↑↑↑↑	↑
Sensitivity to BTK inhibitors	↑↑↑	↑↑	↑	↓
Incidence, %	~60	27-40	27-40	< 10

S338X

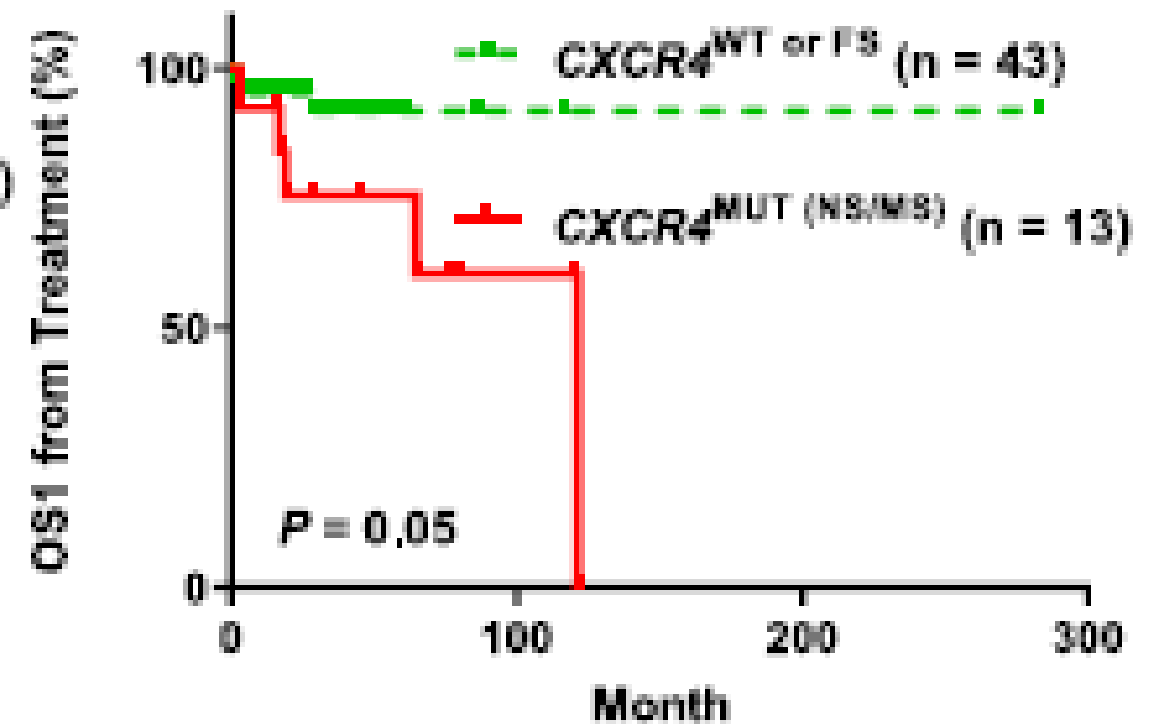
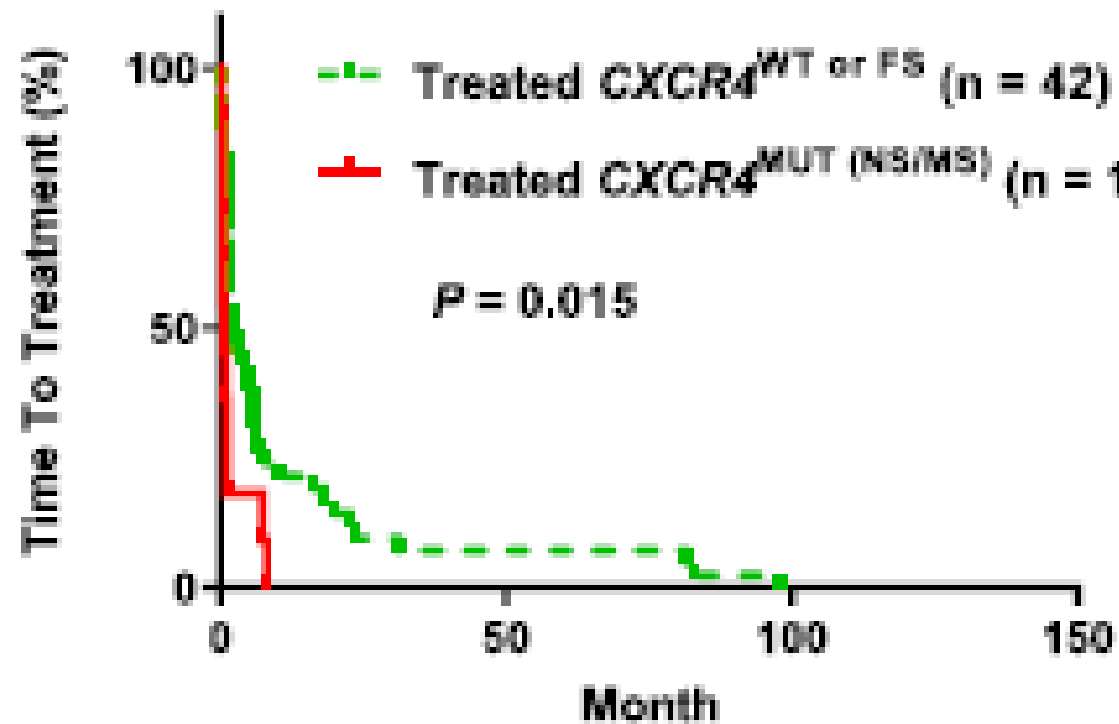
Patients with MYD88 and Nonsense CXCR4 mutations (S338X) show high IGM levels, symptomatic hyperviscosity, and shorter time to initial treatment.

BTK; Bruton's tyrosine kinase

Treon et al, Blood 2014; Schmidt et al, BJH 2015; Abeykoon J, et al. Cancer Manage and Res. 2017;9:73-83; Wang et al, Neoplasia 2021.

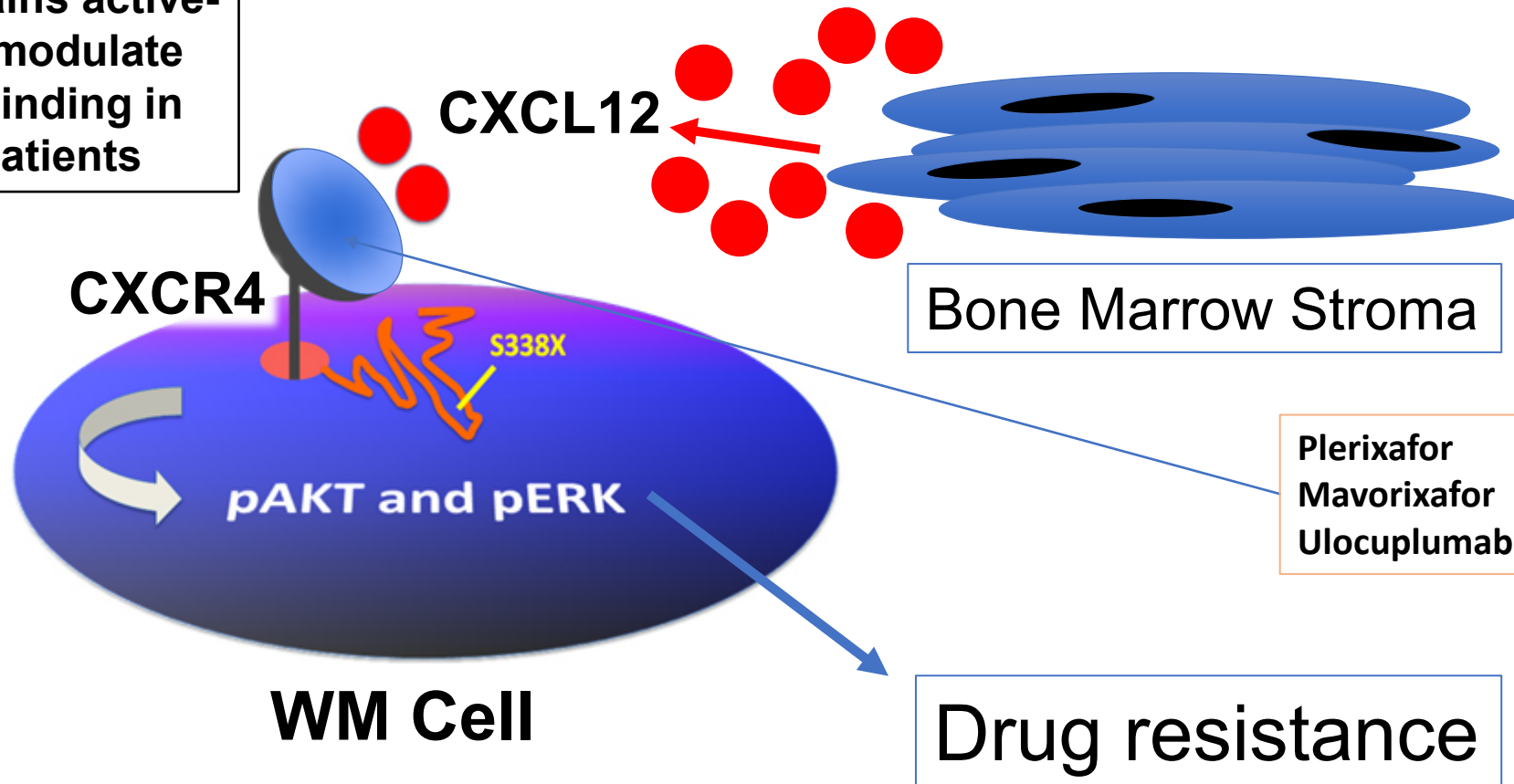
Time to Treatment and OS based on CXCR4 mutation status in WM/LPL patients

MD Anderson Study



Mutated CXCR4 permits ongoing pro-survival signaling by CXCL12

CXCR4 receptor remains active and does not downmodulate following CXCL12 binding in CXCR4 mutated patients



30-40% of WM patients have mutations in CXCR4

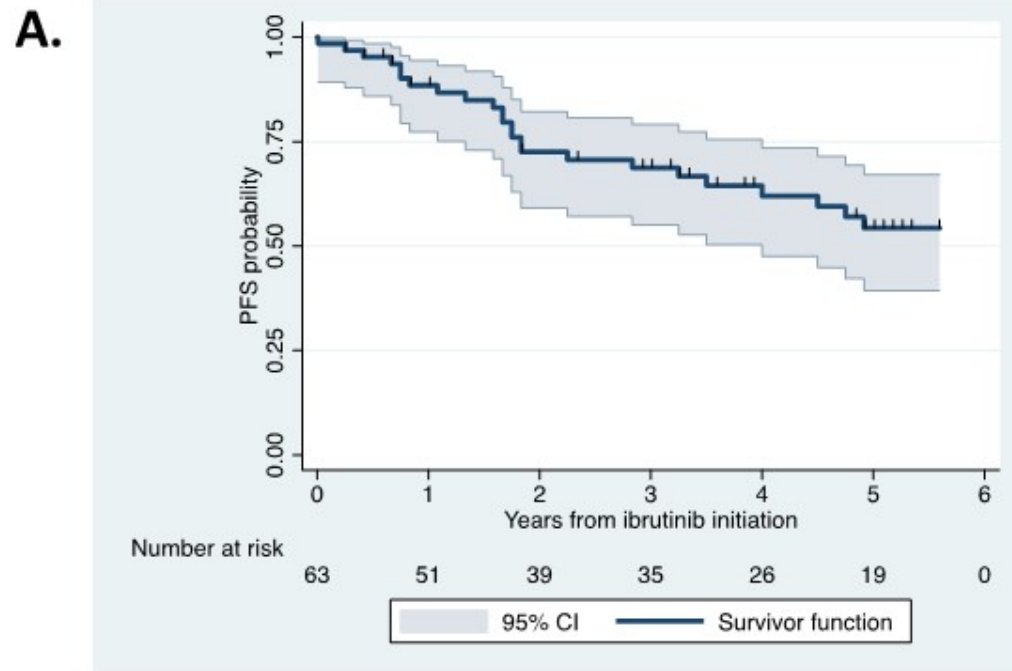
Ibrutinib Activity in Previously Treated WM: Update of the Pivotal Trial (median f/u 59 mos)

	All Patients	MYD88 ^{MUT} CXCR4 ^{WT}	MYD88 ^{MUT} CXCR4 ^{MUT}	MYD88 ^{WT} CXCR4 ^{WT}	P-value
N=	63	36	22	4	N/A
Overall Response Rate-no. (%)	90.5%	100%	86.4%	50%	<0.01
Major Response Rate-no. (%)	79.4%	97.2%	68.2%	0%	<0.0001
Categorical responses					
Minor responses-no. (%)	11.1%	2.8%	18.2%	50%	<0.01
Partial responses-no. (%)	49.2%	50%	59.1%	0%	0.03
Very good partial responses-no. (%)	30.2%	47.2%	9.1%	0%	<0.01
Median time to response (months)					
Minor response (≥Minor response)	0.9	0.9	0.9	0.9	0.38
Major response (≥Partial response)	1.8	1.8	4.7	N/A	0.02

*One patient had MYD88 mutation, but no CXCR4 determination and had SD.

Ibrutinib in Previously Treated WM: Updated PFS

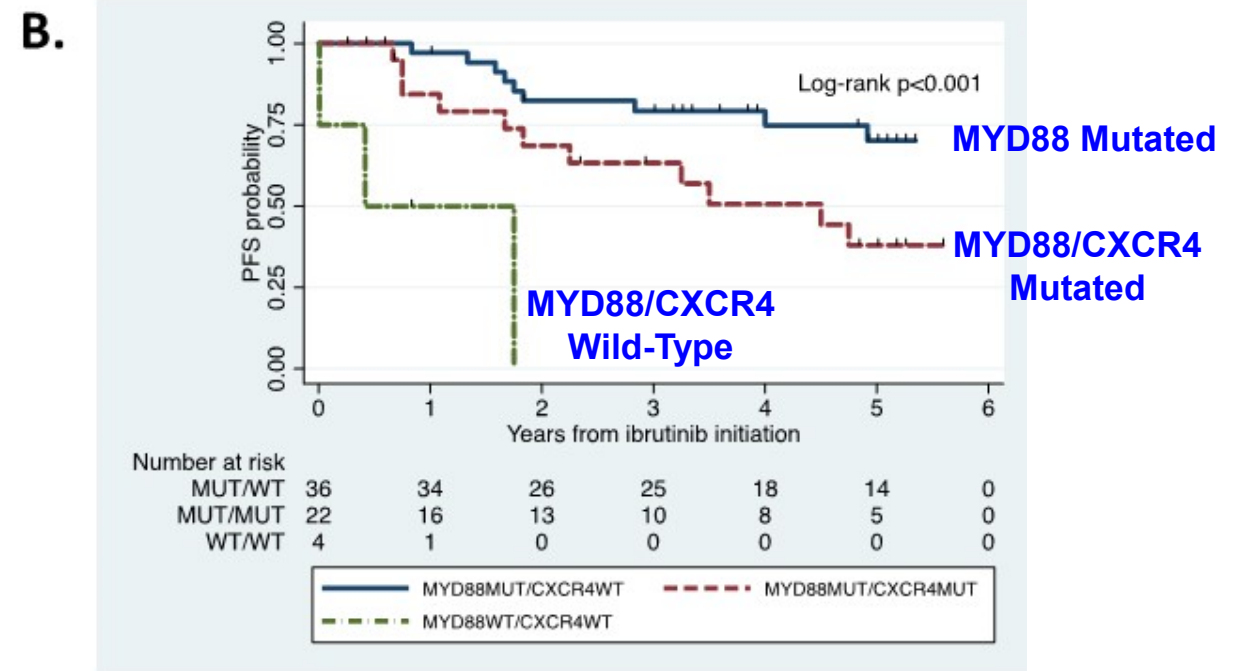
All patients



5 year PFS: 54%

5 year OS: 87%

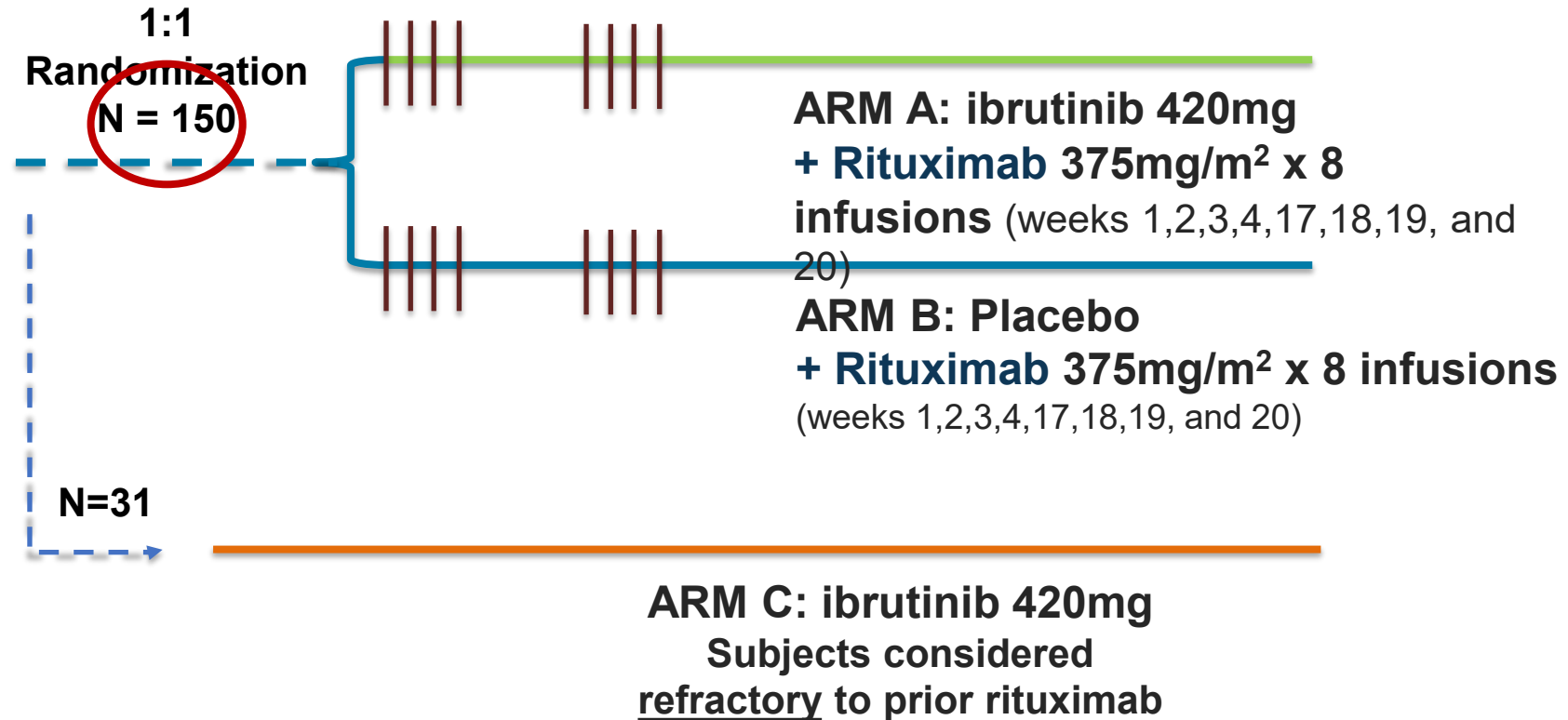
MYD88 and CXCR4 Mutation Status



Treon et al, NEJM 2015; Updated JCO 2021

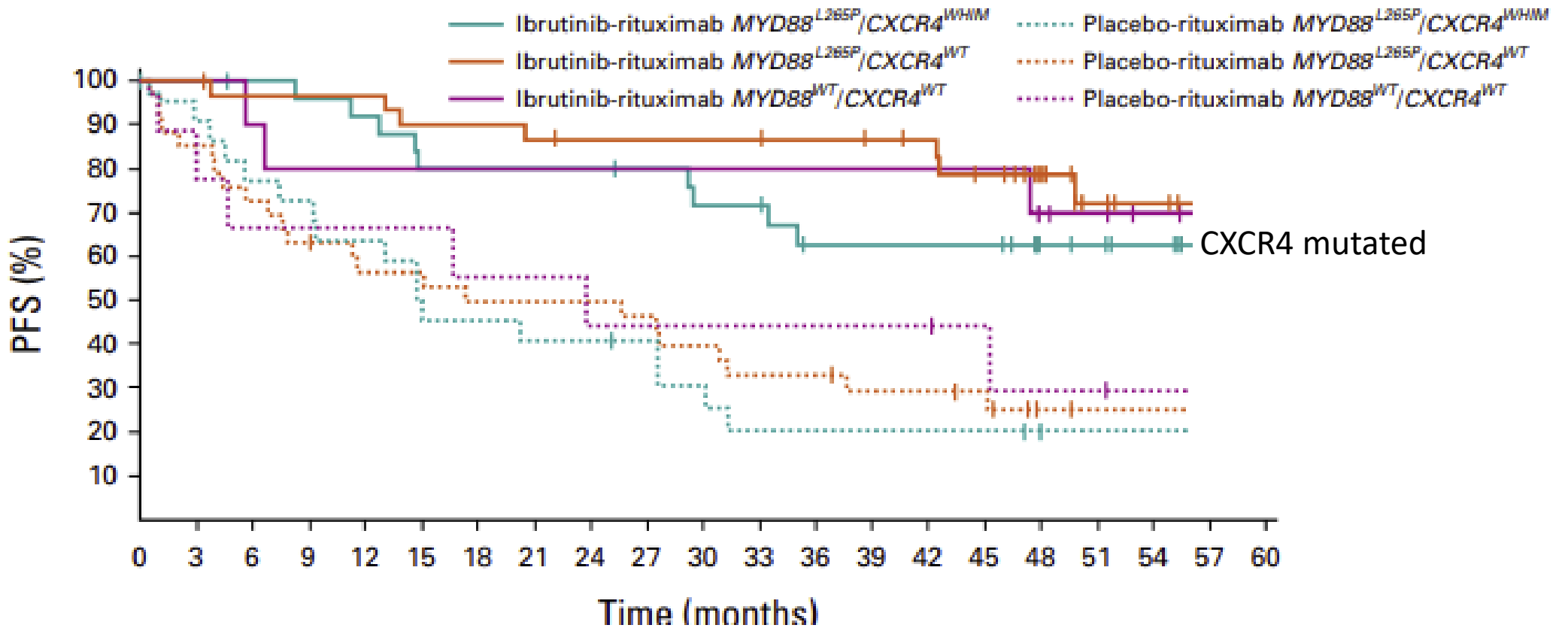
iNNOVATE Study in WM

Treatment Naïve + Previously Treated
45 centers in 9 countries



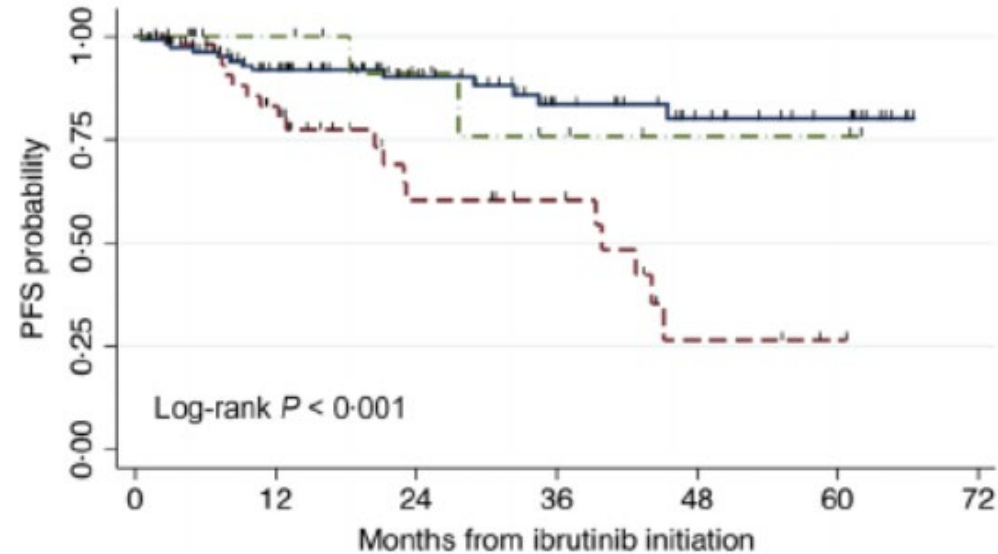
ABC patients genotyped for MYD88 and CXCR4

Progression-Free Survival Benefit: Impact of MYD88/CXCR4 Genotype



CXCR4 Nonsense variants with high clonality impact ibrutinib PFS outcomes

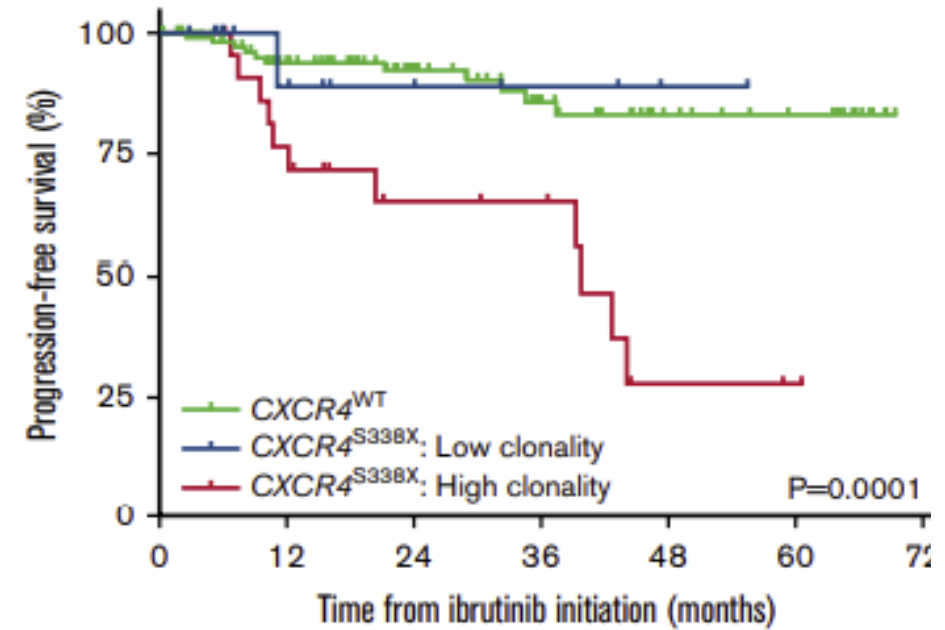
NS vs. FS CXCR4 Mutations



Number at risk	0	12	24	36	48	60	72
CXCR4 WT	112	78	49	30	20	12	0
CXCR4 NS	49	31	14	11	3	1	0
CXCR4 FS	19	13	7	4	2	2	0



High vs. Low NS Clonality CXCR4 Mutations



High clonality $\geq 25\%$

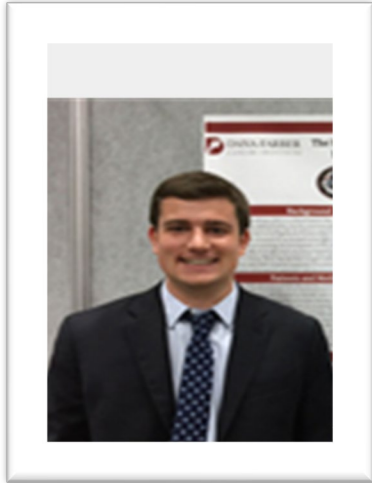


Castillo et al, BJH 2019; Gustine et al, Blood Adv 2019

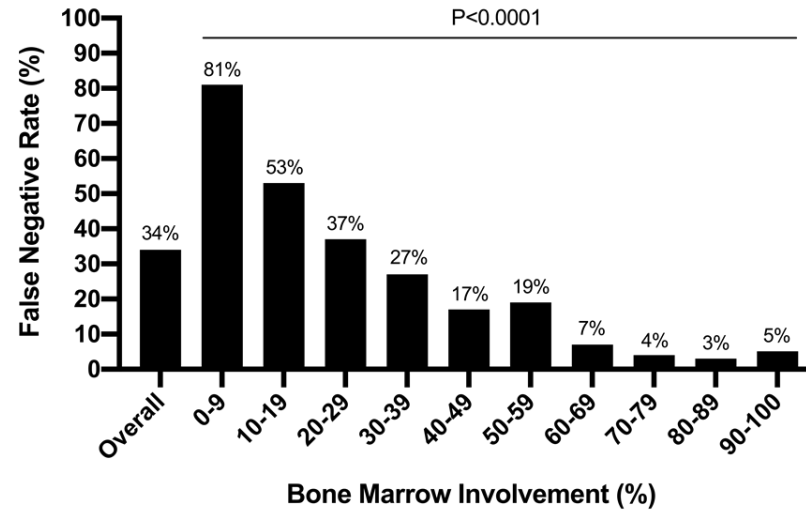
Zanubrutinib: Response by Genotype (ASPEN)

Mutation status	Zanubrutinib (N=101)		Ibrutinib (N=98)	
	MRR	VGPR	MRR	VGPR
Cohort 1				
ALL MYD88 ^{MUT}	77%	28%	78%	19%
MYD88 ^{MUT} CXCR4 ^{WT}	82%	34%	82%	24%
MYD88 ^{MUT} CXCR4 ^{WHIM}	70%	18%	65%	10%

Challenges of CXCR4 detection in WM: Comparison of NGS against AS-PCR/Sanger



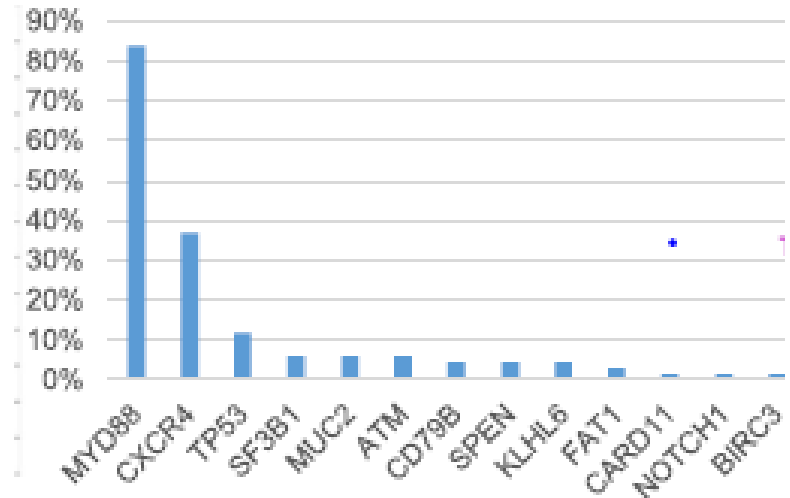
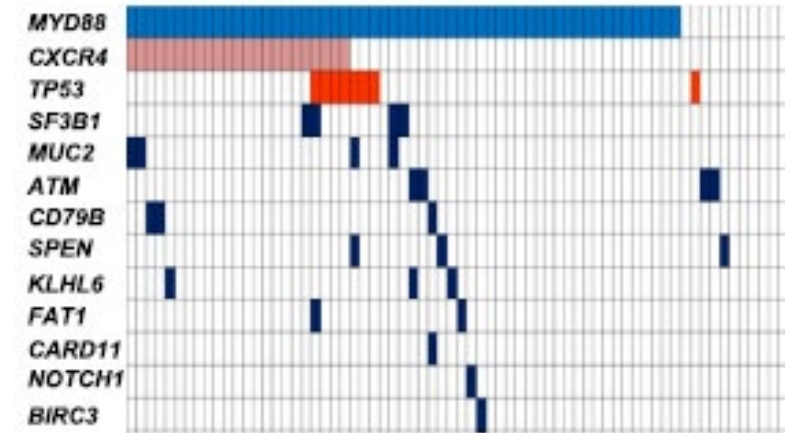
J. Gustine



Sensitivity for mutated CXCR4 detection was 37% by NGS and unselected BM. Low BM involvement and clonality impacted detection.

MD Anderson Sequencing Approach for MYD88 and CXCR4 mutations in WM

Routine clinical workup performed by the CLIA- certified molecular diagnostic laboratory in UT MD Anderson Cancer Center. *MYD88* L265P was determined by AS-PCR and *CXCR4* mutation status (codons 291-353) by targeted NGS. For targeted NGS, genomic DNA extracted from the bone marrow aspirate was used for preparing **sequencing libraries with molecular barcodes** using the Agilent HaloPlex Target Enrichment System (Agilent Technologies), followed by **bidirectional paired-end sequencing using the MiSeq sequencer** (Illumina Inc.). Illumina Experiment Manager, MiSeq Control Software, Real Time Analysis, Sequence Analysis Viewer, MiSeq Reporter, and Agilent SureCall were utilized for experimental setup and NGS data analysis. Although the NGS assay is capable of achieving sensitivity of 1%, the effective lower limit of detection of the assays used for clinical workup was determined to be 5% to 10% taking into consideration the depth of coverage and the ability to confirm low-level mutations using independent conventional platforms.



Cell-free DNA analysis for *MYD88*^{L265P} and *CXCR4*^{S338X} mutations in Waldenström macroglobulinemia.

Variable	<i>MYD88</i> ^{L265P}		<i>CXCR4</i> ^{S338X}	
	BM	cfDNA	BM	cfDNA
True positive – no.	25	20	6	4
True negative – no.	3	3	17	17
False positive – no.	0	0	0	0
False negative – no.	0	5	0	2
Concordance (Cohen κ) – %	Ref.	82 (0.46)	Ref.	91 (0.75)
Sensitivity (95% CI) – %	Ref.	80 (59-92)	Ref.	67 (24-94)
Specificity (95% CI) – %	Ref.	100 (31-100)	Ref.	100 (77-100)
PPV (95% CI) – %	Ref.	100 (80-100)	Ref.	100 (40-100)
NPV (95% CI) – %	Ref.	38 (10-74)	Ref.	89 (65-98)

Adjusted test performance findings for cfDNA using both BM19+ and BM19- fractions as reference tissue.

Demos et al, AJH 2021



NCCN Guidelines Version 1.2022

Waldenström Macroglobulinemia/ Lymphoplasmacytic Lymphoma



NCCN Categories of Evidence and Consensus	
Category 1	Based upon high-level evidence, there is uniform NCCN consensus that the intervention is appropriate.
Category 2A	Based upon lower-level evidence, there is uniform NCCN consensus that the intervention is appropriate.
Category 2B	Based upon lower-level evidence, there is NCCN consensus that the intervention is appropriate.
Category 3	Based upon any level of evidence, there is major NCCN disagreement that the intervention is appropriate.

All recommendations are category 2A unless otherwise indicated.

NCCN Categories of Preference	
Preferred intervention	Interventions that are based on superior efficacy, safety, and evidence; and, when appropriate, affordability.
Other recommended intervention	Other interventions that may be somewhat less efficacious, more toxic, or based on less mature data; or significantly less affordable for similar outcomes.
Useful in certain circumstances	Other interventions that may be used for selected patient populations (defined with recommendation).

All recommendations are considered appropriate.

- Category 1 studies are generally supported by large randomized trials
- Category 2 studies are generally supported by prospective Phase II trials. Larger multicenter studies, with confirmed findings generally rate 2A.
- Category 3 studies may include small patient series, or retrospective data analysis.



NCCN Guidelines Version 1.2022

Waldenström Macroglobulinemia/ Lymphoplasmacytic Lymphoma



PRIMARY THERAPY FOR WM/LPL ^a (Order of regimens is alphabetical and does not indicate preference)	
Preferred Regimens	
<ul style="list-style-type: none"> • Bendamustine/rituximab • Bortezomib/dexamethasone/rituximab^b • Ibrutinib ± rituximab (category 1) 	<ul style="list-style-type: none"> • Rituximab/cyclophosphamide/dexamethasone • Zanubrutinib (category 1)
Other Recommended Regimens	
<ul style="list-style-type: none"> • Bendamustine • Bortezomib ± rituximab^b • Bortezomib/dexamethasone • Carfilzomib/rituximab/dexamethasone • Cladribine ± rituximab^c 	<ul style="list-style-type: none"> • Fludarabine ± rituximab^c • Fludarabine/cyclophosphamide/rituximab^c • Ixazomib/rituximab/dexamethasone • Rituximab • Rituximab/cyclophosphamide/prednisone

^a See [General Considerations for Systemic Therapy for WM/LPL \(WM/LPL-B 1 of 4\)](#).

^b Consider for patients presenting with symptomatic hyperviscosity, or in whom rapid IgM reduction is required.

^c May be associated with disease transformation and/or development of MDS/AML in patients with Waldenström macroglobulinemia.

Note: All recommendations are category 2A unless otherwise indicated.

Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

[Continued
References](#)

WM/LPL-B
2 OF 4

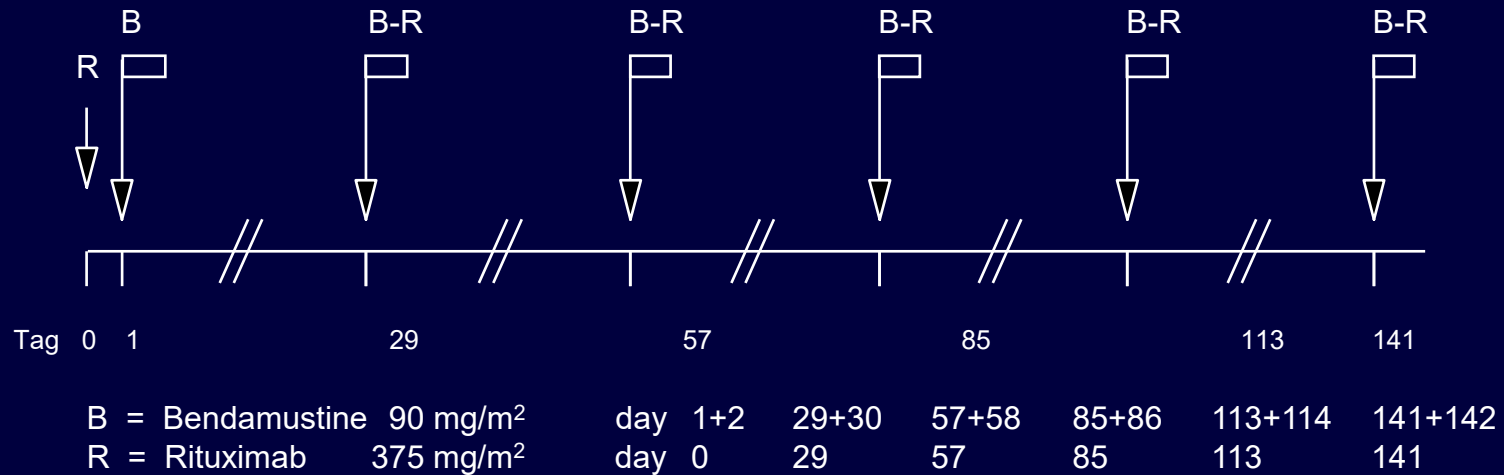
STIL GROUP STUDY: Bendamustine-Rituximab vs. CHOP-R



M. Rummel

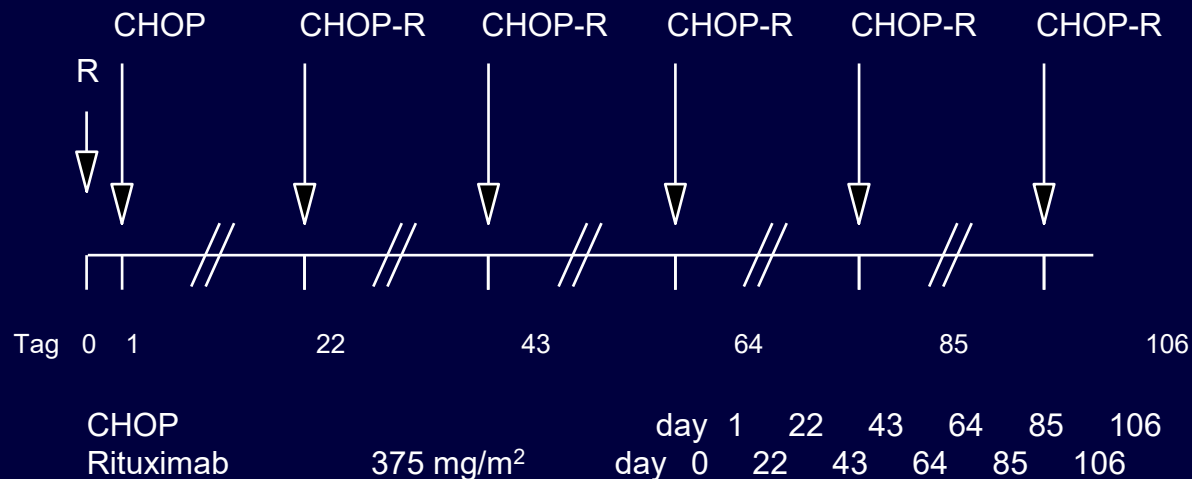
N=549

Bendamustine plus Rituximab (B-R)



Randomization

CHOP plus Rituximab (CHOP-R)

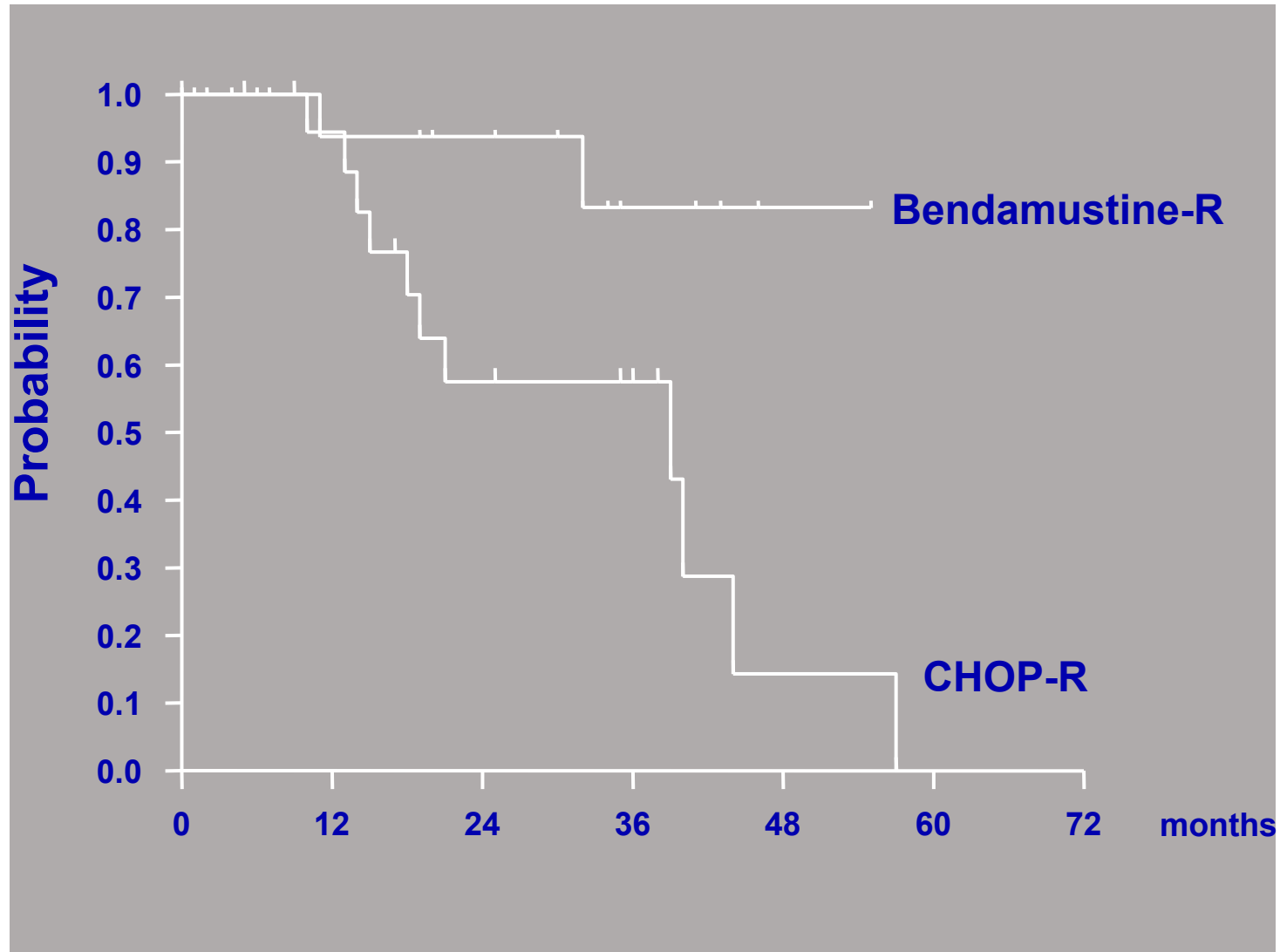


Waldenström patients characteristics: B-R vs CHOP-R

N=40

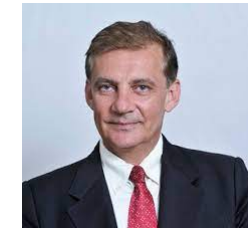
20 th Aug 2008 40 evaluable pts.	Bendamustine-R (n=23)	CHOP-R (n=17)
Age (median)	65 yrs	64 yrs
Stage IV	100 %	100 %
Beta-2 Microglobulin	3,2	3,4
IgM (median, mg/dl)	2.790	1.690
Range IgM (mg/dl)	11.220 - 1.100	8.510 - 900
Hb (median, g/dl)	10,2	9,9
Toxicity treatment associated		
Neuropathy (patients)	1	3

PFS: Benda-R vs CHOP-R in frontline WM

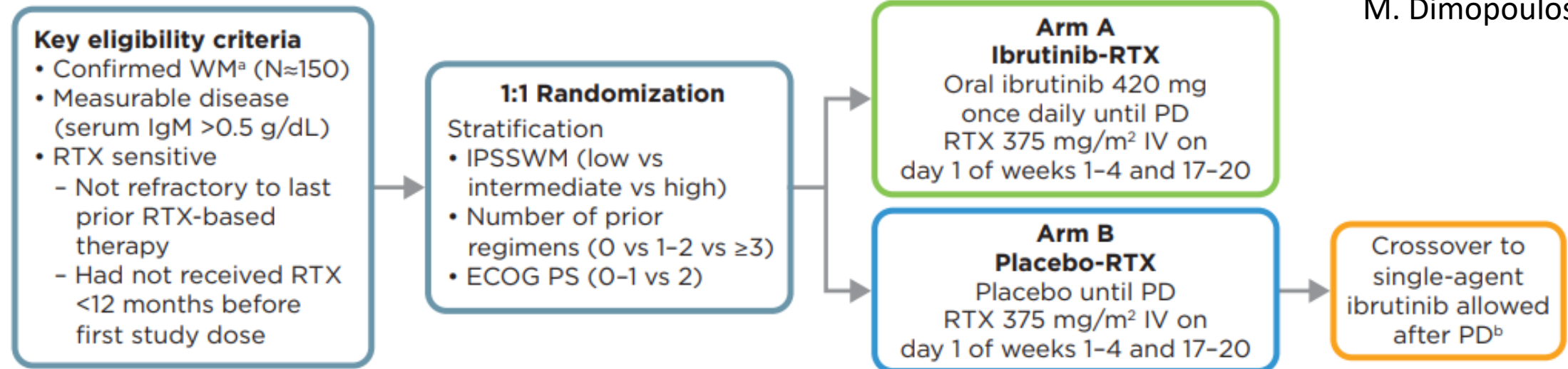


iINNOVATE Study Design

C. Buske



M. Dimopoulos



ECOG PS, Eastern Cooperative Oncology Group performance status; IPSSWM, International Prognostic Scoring System for Waldenström's Macroglobulinemia; IRC, independent review committee; IV, intravenous; PD, progressive disease.

^aPreviously untreated patients were allowed to enroll following a protocol amendment (November 2015); therefore, their enrollment started later than patients who had relapsed.

^bPatients in the placebo-RTX arm could receive next-line single-agent ibrutinib in crossover following IRC-confirmed PD.

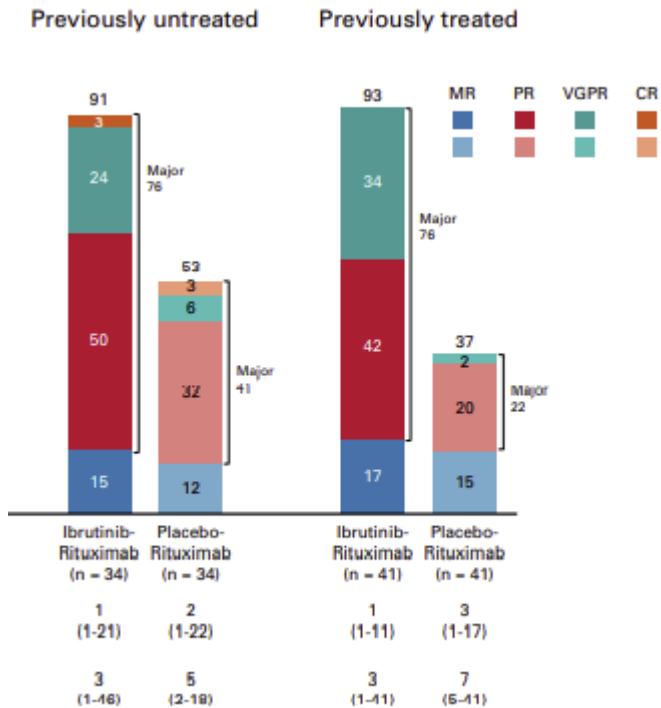
- iINNOVATE (PCYC-1127) was a double-blind, randomized, placebo-controlled, multicenter, international phase 3 study to assess the efficacy and safety of ibrutinib-RTX versus placebo-RTX in patients with WM (**Figure 1**).
- The primary endpoint was PFS by IRC. Secondary endpoints included response rate by IRC, time to next treatment, hemoglobin (Hgb) improvement, overall survival (OS), and safety.
- After study closure, patients without PD could continue ibrutinib in an extension program.

iNNOVATE: Baseline Characteristics

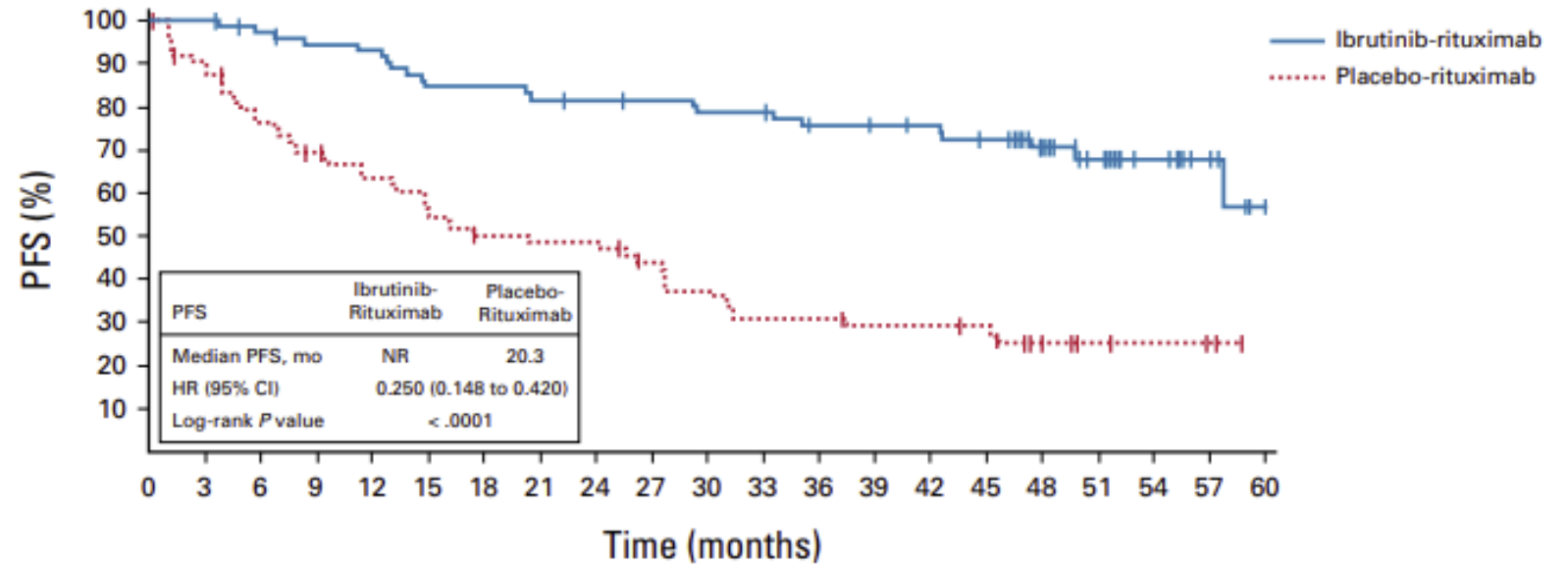
Characteristic	Ibrutinib + R (n=75)	Placebo + R (n=75)
Age, median (range), y	70 (36-89)	68 (39-85)
Male sex, n (%)	45 (60)	54 (72)
IPSSWM, n (%)		
Low	15 (20)	17 (23)
Intermediate	33 (44)	28 (37)
High	27 (36)	30 (40)
Medium serum IgM, g/L (range)	33 (6-78)	32 (6-83)
Number of prior systemic therapies, n (%)		
0	34 (45)	34 (45)
1-2	34 (45)	36 (48)
≥3	7 (9)	5 (7)
Genotype, n (%)		
<i>MYD88</i> ^{L265P} / <i>CXCR4</i> ^{WT}	32 (43)	35 (47)
<i>MYD88</i> ^{L265P} / <i>CXCR4</i> ^{WT}	26 (35)	23 (31)
<i>MYD88</i> ^{L265P} / <i>CXCR4</i> ^{WT}	11 (15)	9 (12)
Unkown	6 (8)	8 (11)
Bone marrow infiltration: mean % cellularity (range)	73 (25-100)	75 (2-100)

iNNOVATE Final Analysis

Response Rates



Progression-Free Survival





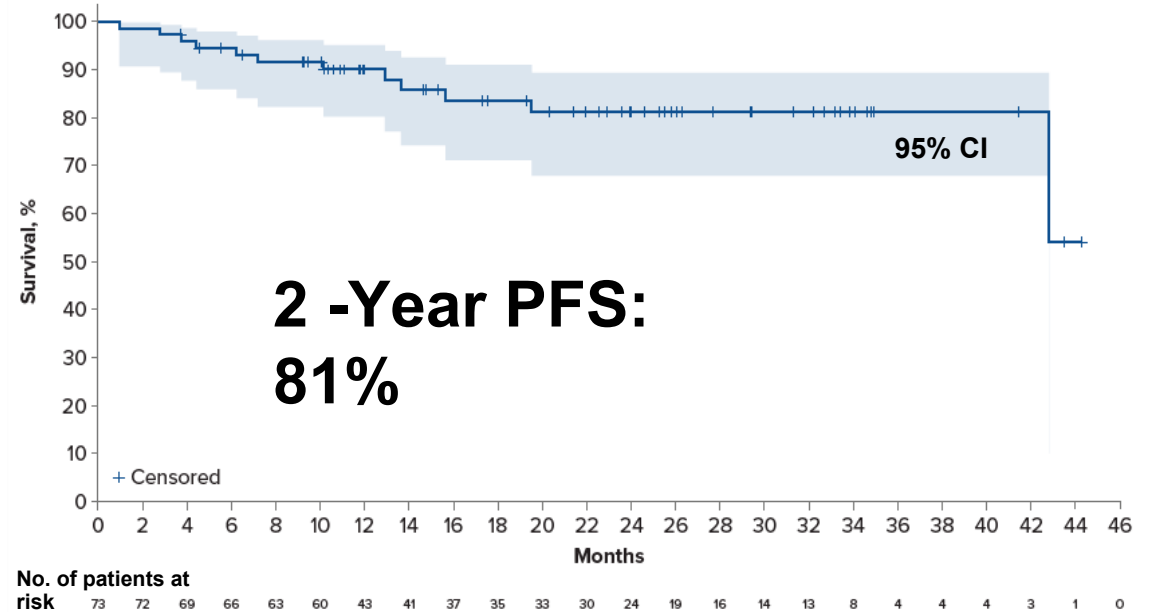
Zanubrutinib in WM:

Phase 2 data in TN and previously treated pts.

Best Response in WM	zanubrutinib		
	Overall	TN	RR
Evaluable for efficacy, n	73	24	49
Median Follow-up	23.9 mo	12.3 mo	24.8 mo
Response Criteria	Mod. 6 th IWWM (IgM decreases only, and not extramedullary disease)		
Median Prior Lines of Therapy		0	2 (1-8)
ORR	92%	96%	90%
MRR	82%	87%	78%
CR/VGPR ¹	42%	29%	49%
PR	40%	58%	31%

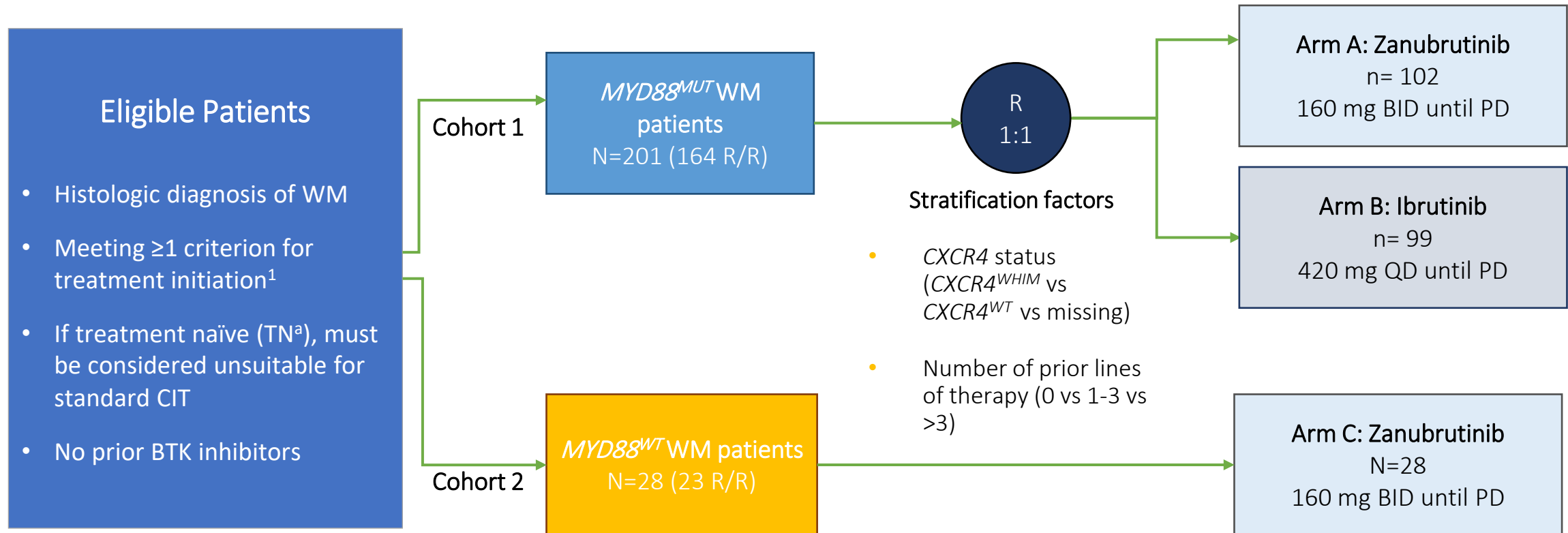
Zanubrutinib 320 mg qd to 160 mg BID

Progression Free Survival (PFS)



Trotman et al, EHA 2019; Blood 2020;
10.1182/blood.2020006449. Online ahead of print.

ASPEN: A Phase 3 Study of Zanubrutinib vs. Ibrutinib in WM



BID, twice daily; BTK, Bruton tyrosine kinase; CIT, chemoimmunotherapy; *CXCR4*, C-X-C Motif Chemokine Receptor 4; *MYD88^{MUT}*, myeloid differentiation primary response gene 88 mutant; PD, progressive disease; QD, daily; R, randomization; R/R, relapsed/refractory; TN, treatment naïve; WM, Waldenström Macroglobulinemia; WT, wild-type.

^aUp to 20% of the overall population.

1. Dimopoulos MA, et al. *Blood*. 2014;124:1404-1411.

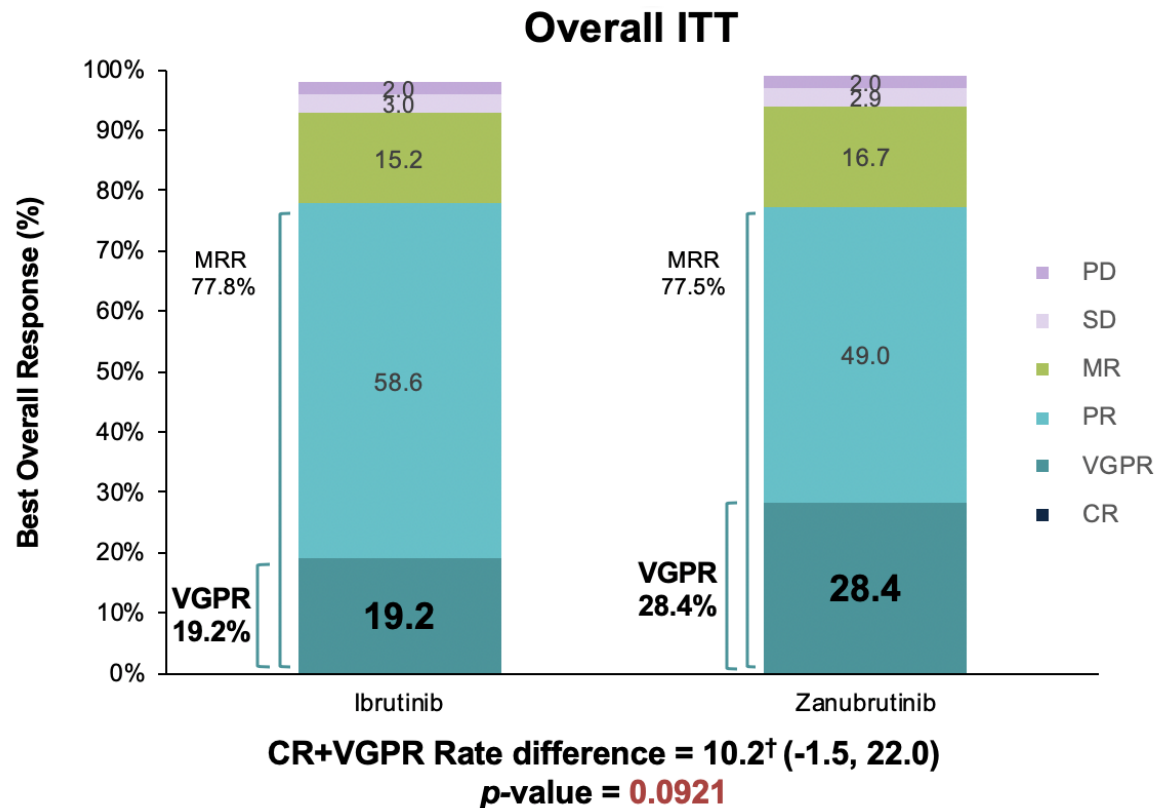
ASPEN Cohort 1: Demographics and Disease Characteristics

Characteristics, n (%)	Ibrutinib (n = 99)	Zanubrutinib (n =102)
Age, years median (range) > 75 years	70.0 (38, 90) 22 (22)	70.0 (45, 87) 34 (33)
Gender, n (%) Male	65 (66)	69 (68)
Prior lines of therapy, n (%) 0 1-3 >3	18 (18) 74 (75) 7 (7)	19 (19) 76 (75) 7 (7)
Genotype by central lab, n (%)* MYD88 ^{L265P} /CXCR4 ^{WT} MYD88 ^{L265P} /CXCR4 ^{WHIM}	90 (91) 8 (8)	91 (89) 11 (11)
IPSS WM ¹ Low Intermediate High	13 (13) 42 (42) 44 (44)	17 (17) 38 (37) 47 (46)
Hemoglobin ≤ 110 g/L	53 (54)	67 (66)

*"Wildtype-blocking PCR" for MYD88 and Sanger sequencing for CXCR4 using bone marrow aspirates. One patient had local NGS testing results of MYD88 L265P/ CXCR4 Unknown.

Tam CS, et al. *Blood*. 2020. Online ahead of print.

ASPEN Cohort 1: Efficacy, Response by IRC



- Data cutoff: August 31, 2019
- Superiority in CR+VGPR rate compared to ibrutinib in relapsed/refractory population (primary study hypothesis) was not significant* (p-value 0.1160)

CR, complete response; IRC, independent review committee; ITT, intention-to-treat; MRR, major response rate; MR, minor response; PD, progressive disease; PR, partial response; SD, stable disease; VGPR, very good PR.

Overall concordance between Independent review and investigators = 94%

*All other P values are for descriptive purposes only. [†]Adjusted for stratification factors and age group.

1. Tam CS et al. *J Clin Oncol.* 2020;38(15 Suppl):8007.

ASPEN: AE Categories of Interest (BTKi Class AEs)

AE Categories, n (%) (pooled terms)	All Grades		Grade ≥ 3	
	Ibrutinib (n = 98)	Zanubrutinib (n = 101)	Ibrutinib (n = 98)	Zanubrutinib (n = 101)
Atrial fibrillation/ flutter [†]	15 (15.3)	2 (2.0)	4 (4.1)	0 (0.0)
Diarrhea (PT)	31 (31.6)	21 (20.8)	1 (1.0)	3 (3.0)
Hemorrhage	58 (59.2)	49 (48.5)	8 (8.2)	6 (5.9)
Major hemorrhage ^a	9 (9.2)	6 (5.9)	8 (8.2)	6 (5.9)
Hypertension	17 (17.3)	11 (10.9)	12 (12.2)	6 (5.9)
Neutropenia ^{b†}	13 (13.3)	30 (29.7)	8 (8.2)	20 (19.8)
Infection	66 (67.3)	67 (66.3)	19 (19.4)	18 (17.8)
Second Malignancy	11 (11.2)	12 (11.9)	1 (1.0)	2 (2.0)

Higher AE rate in bold blue with ≥ 10% difference in any grade or ≥ 5% difference in grade 3 or above.

No tumor lysis syndrome was reported. Opportunistic infection ibrutinib (n=2), zanubrutinib (n=1).

AE, adverse event; BTKi, Bruton tyrosine kinase inhibitor; PT, preferred term.

^aDefined as any grade ≥ 3 hemorrhage or any grade central nervous system hemorrhage.

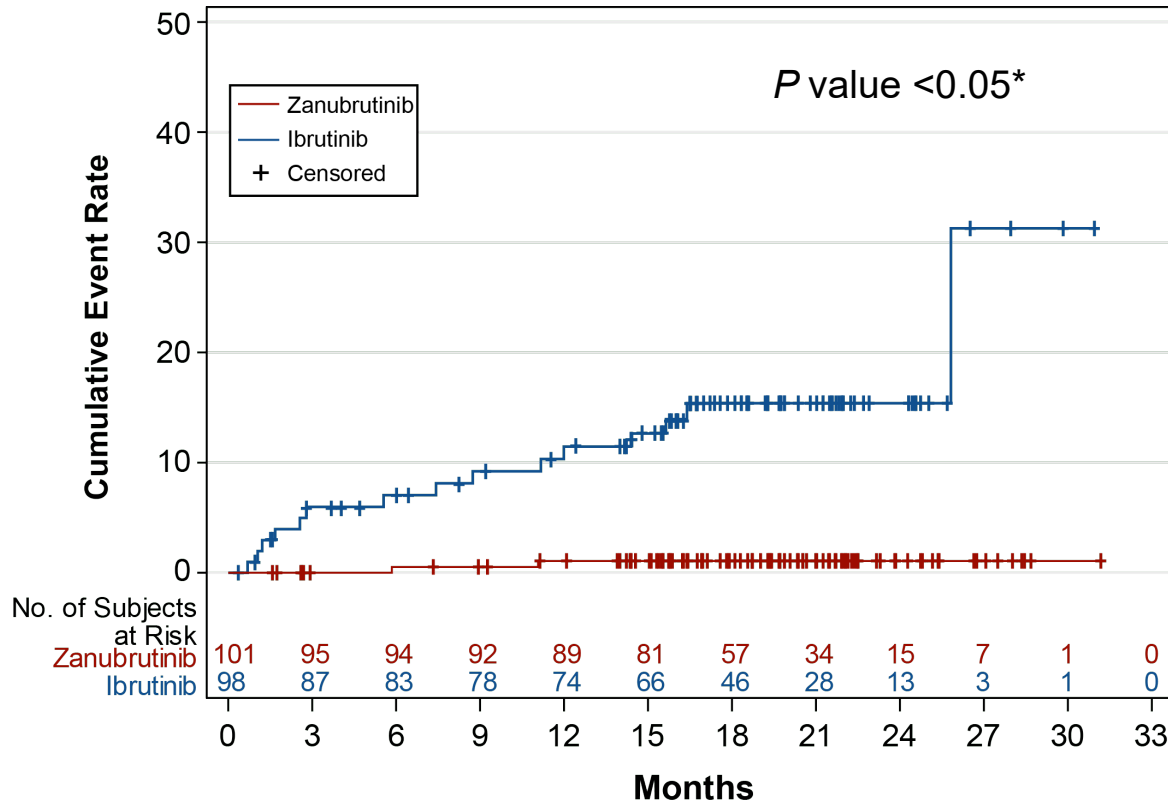
^bIncluding PT terms of neutropenia, neutrophil count decreased, febrile neutropenia, agranulocytosis, neutropenic infection and neutropenic sepsis.

[†] Descriptive two-sided *P*-value < 0.05.

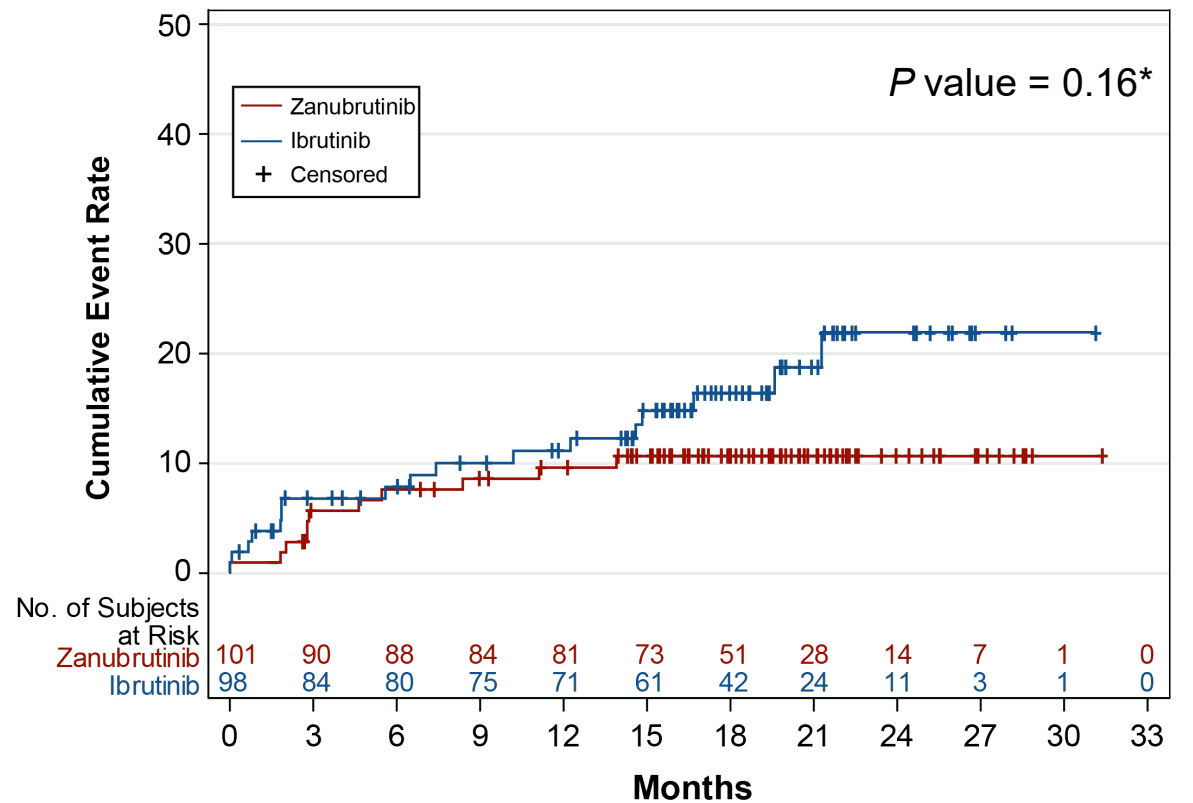
Tam et al, Blood 2020

ASPEN Cohort 1: Time to AE, Risk Analysis Over Duration of Treatment

Kaplan-Meier Curve: Time to **Atrial Fibrillation/Flutter**



Kaplan-Meier Curve: Time to **Hypertension**



AE, adverse event.
 *Descriptive purpose only.
 Tam CS et al. *J Clin Oncol*. 2020;38(15 Suppl):8007.

ASPEN Cohort 2: Patient and Disease Characteristics

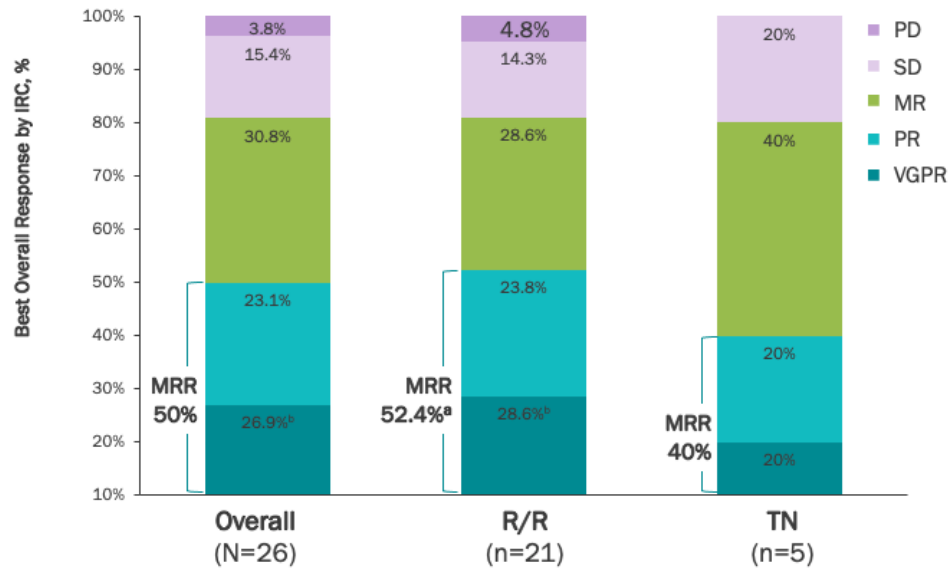
Characteristic	Total (N=28)
Age, median (range), y	70.1 (39-87)
>65 years, n (%)	19 (67.9)
>75 years, n (%)	12 (42.9)
Male sex, n (%)	14 (50)
IPSSWM, n (%)	
Low	5 (17.9)
Intermediate	11 (39.3)
High	12 (42.9)
Prior treatment status	
Treatment-naïve, n (%)	5 (17.9)
R/R, n (%)	23 (82.1)
No. of prior therapies for R/R pts, median (range)	1 (1-5)
Extramedullary disease present at baseline by IRC, n (%)	21 (75.0)
Median bone marrow involvement (n=26, central identified assay)	23%
Median bone marrow involvement (n=24, 2 pts with MYD88 ^{L265P} by NGS)	15%
Bone marrow involvement >25%, (n=24)	11

IPSSWM, International Prognostic Scoring System Waldenström Macroglobulinemia; pt, patient; R/R, relapsed/refractory; IRC, independent review committee.

Dimopoulos MA, et al. EHA 2020. EP1180.

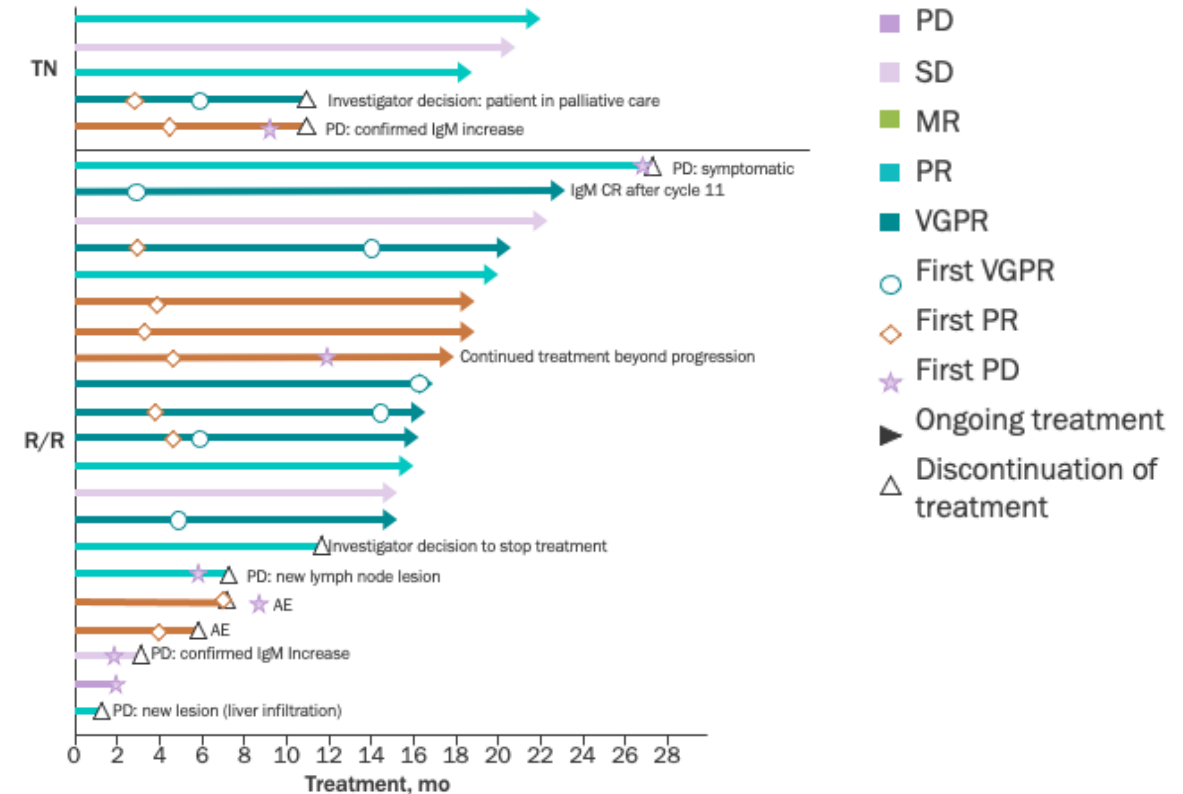
ASPEN Cohort 2: Response

Response by IRC



- Major response rate of 50.0% including 26.9% with VGPR
- Median time to first major response (partial response or better, requiring reduction in extramedullary disease if present at baseline), was 2.9 months (range, 1.9-16.1)
- Of the 11 patients with median BM involvement >25%: 3 VGPR, 5 PR, 2 MR, 1 SD, MRR=72.7%

Response Over Time on Treatment



- Color of bars represents the best response for each patient

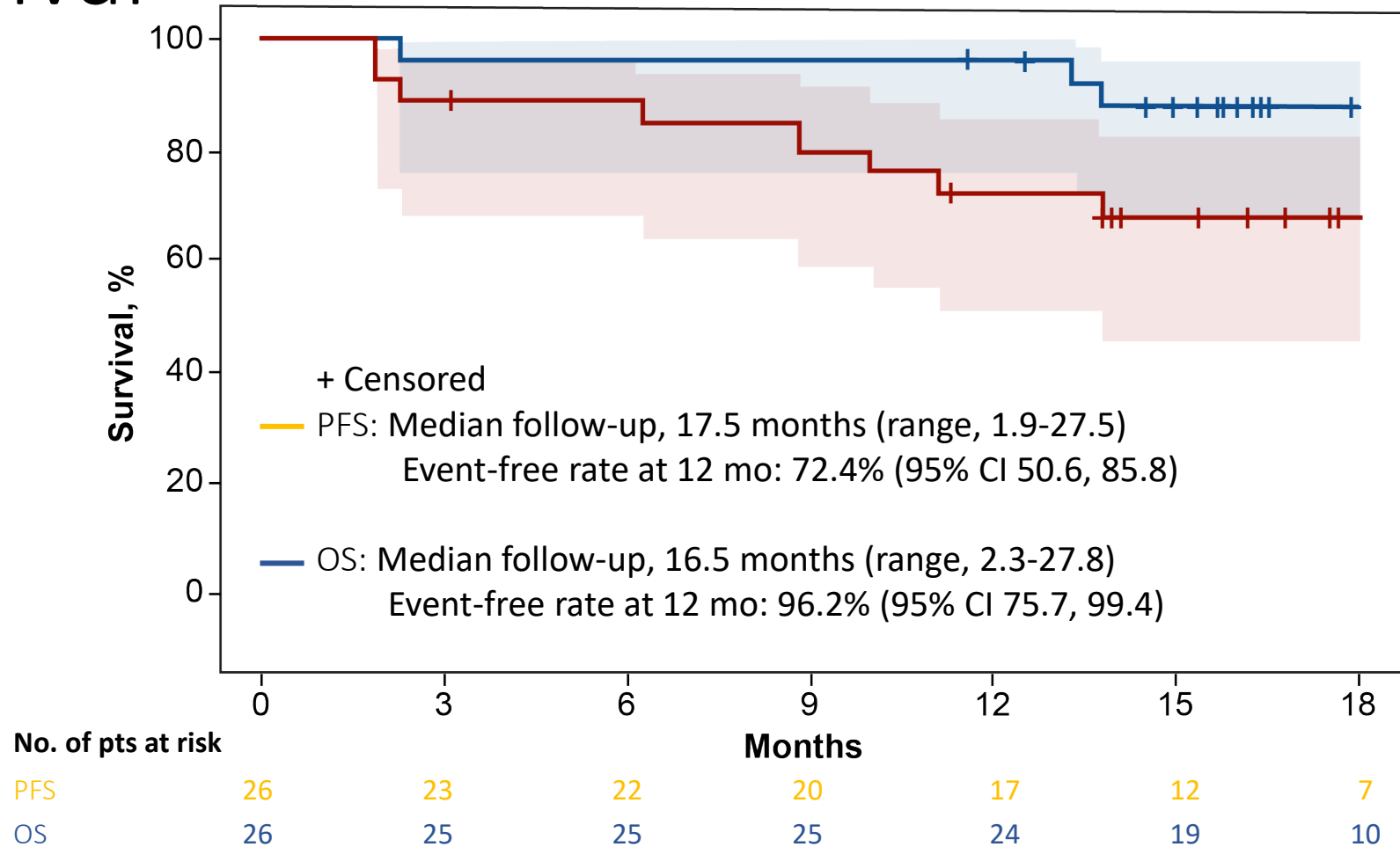
Data cutoff: August 31, 2019

AE, adverse event; CR, complete response; IgM, immunoglobulin M, IRC, independent review committee; MR, minor response; MRR, major response rate (≥PR); PD, progressive disease; PR, partial response; pt, patient; R/R, relapsed/refractory; SD, stable disease; TN, treatment-naïve; VGPR, very good PR.

^aIncluding pts confirmed by next-generation sequencing of no other activating *MYD88* mutations. ^bOne pt achieved IgM complete response (normalized IgM and negative immunofixation since cycle 11, with bulky extramedullary disease improving).

Dimopoulos MA, et al. EHA 2020. EP1180.

ASPEN Cohort 2: Progression-Free and Overall Survival



Shaded areas show the 95% CI.
 OS, overall survival; PFS, progression-free survival; pt, patient.
 Dimopoulos MA, et al. EHA 2020. EP1180.



NCCN Guidelines Version 1.2022

Waldenström Macroglobulinemia/ Lymphoplasmacytic Lymphoma



THERAPY FOR PREVIOUSLY TREATED WM/LPL ^a (Order of regimens is alphabetical and does not indicate preference)	
Preferred Regimens	
<ul style="list-style-type: none"> • Bendamustine/rituximab • Bortezomib/dexamethasone/rituximab^b • Ibrutinib ± rituximab (category 1) 	<ul style="list-style-type: none"> • Rituximab/cyclophosphamide/dexamethasone • Zanubrutinib (category 1)
Other Recommended Regimens	
<ul style="list-style-type: none"> • Acalabrutinib • Bendamustine • Bortezomib ± rituximab^b • Bortezomib/dexamethasone • Cladribine ± rituximab^c 	<ul style="list-style-type: none"> • Cyclophosphamide/doxorubicin/vincristine/prednisone/rituximab • Fludarabine ± rituximab^c • Fludarabine/cyclophosphamide/rituximab^c • Rituximab • Rituximab/cyclophosphamide/prednisone
Useful In Certain Circumstances	
<ul style="list-style-type: none"> • Everolimus • Ofatumumab (for rituximab-intolerant individuals)^d 	
Hematopoietic Cell Transplant	
<ul style="list-style-type: none"> • In selected cases hematopoietic cell transplantation may be appropriate with either: <ul style="list-style-type: none"> ▶ Allogeneic hematopoietic cell transplant (ablative or nonablative)^e ▶ Autologous hematopoietic cell transplant 	

^a See [General Considerations for Systemic Therapy for WM/LPL \(WM/LPL-B 1 of 4\)](#).

^b Consider for patients presenting with symptomatic hyperviscosity, or in whom rapid IgM reduction is required.

^c May be associated with disease transformation and/or development of MDS/AML in patients with Waldenström macroglobulinemia.

^d Ofatumumab may be used for rituximab-intolerant individuals as a single agent or in combination therapy anywhere that rituximab is given. While ofatumumab is no longer commercially available, it may be obtained for clinical use.

^e Should ideally be undertaken in the context of a clinical trial.

Note: All recommendations are category 2A unless otherwise indicated.

Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

References

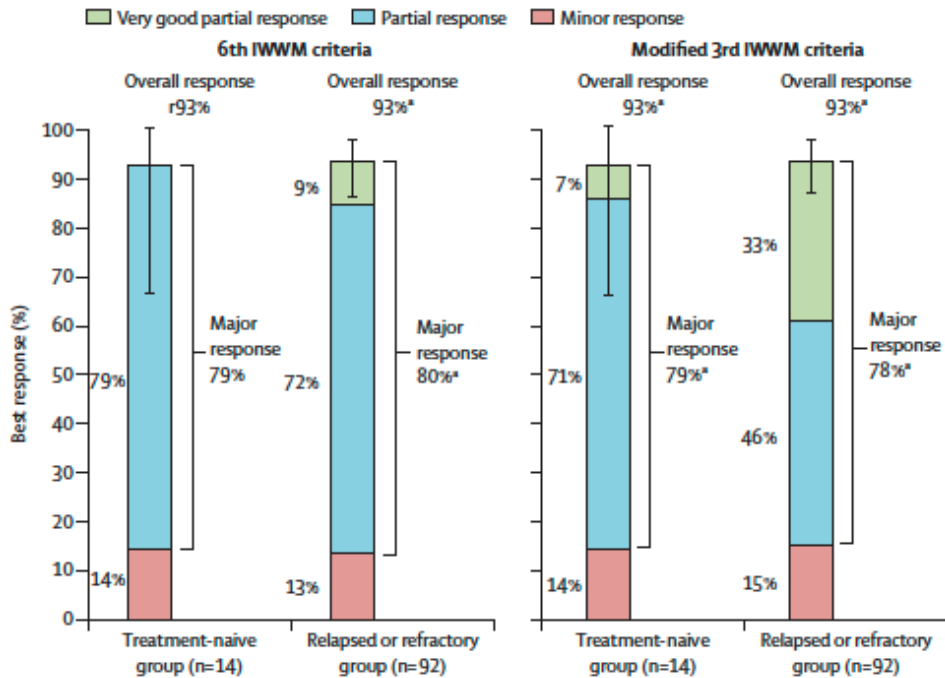
Acalabrutinib Phase 2 WM Study: Baseline Characteristics

Characteristic	Treatment Naïve (n=14)	Relapsed/Refractory (n=92)
Age, median (range), y	73 (48-86)	69 (39-90)
Male sex, n (%)	10 (71%)	63 (68%)
ECOG PS, n (%)		
≤1	12 (86%)	88 (96%)
0	3 (21%)	52 (57%)
1	9 (64%)	36 (39%)
2	2 (14%)	4 (4%)
Median time since initial WM diagnosis (range), y	0.4 (0.04-5.8)	6.1 (0.2-25.4)
Extramedullary disease ^a , n (%)	9 (64)	59 (64)
Median time to last treatment, months (range)	NA	16.2 (0.03-89.6)
Median number of previous therapies (range)	NA	2 (1-7)
≥3 previous therapies	NA	41 (45%)

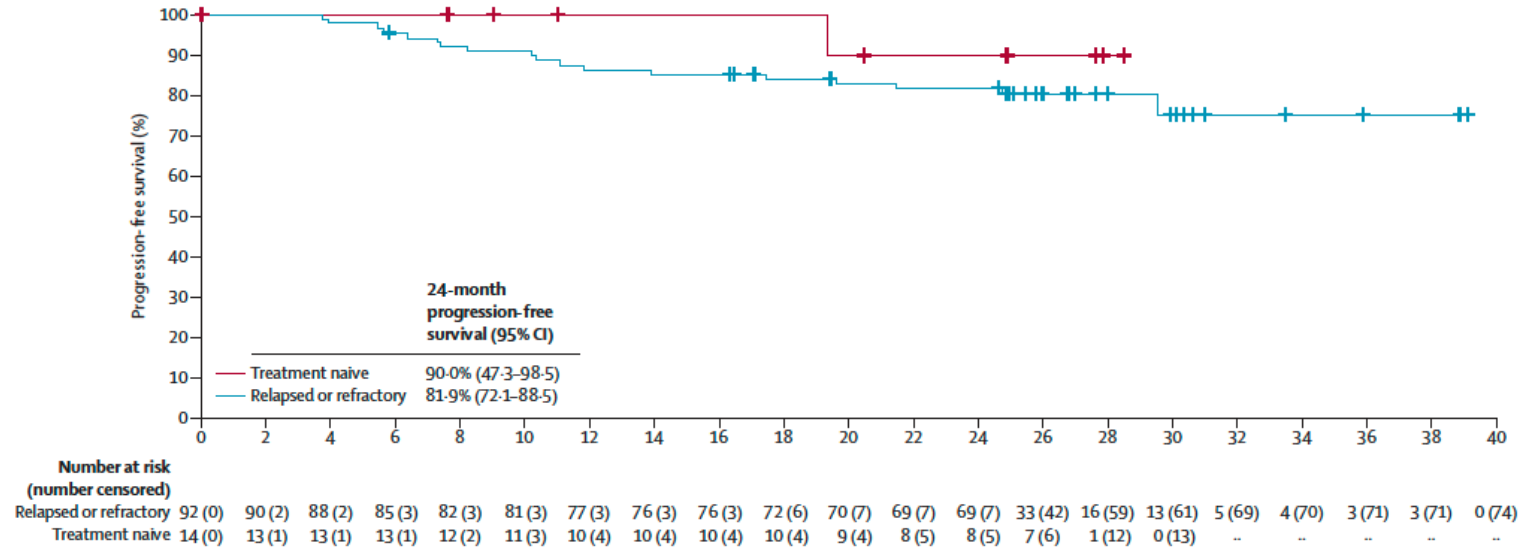
^aDefined as lymphadenopathy (>1.5 cm) and splenomegaly (an enlarged spleen of any size).

Acalabrutinib Phase 2 WM Study: Efficacy

Overall Response



Progression-Free Survival



- Median duration of follow-up was 27.4 months
- Median duration of response has not been reached
 - 24-month duration of response for treatment-naïve patients (90%) and relapsed/refractory patients (82%)
- Overall survival was 92% in treatment-naïve patients and 89% in relapsed/refractory patients

Acalabrutinib Phase 2 WM Study: Safety and Tolerability

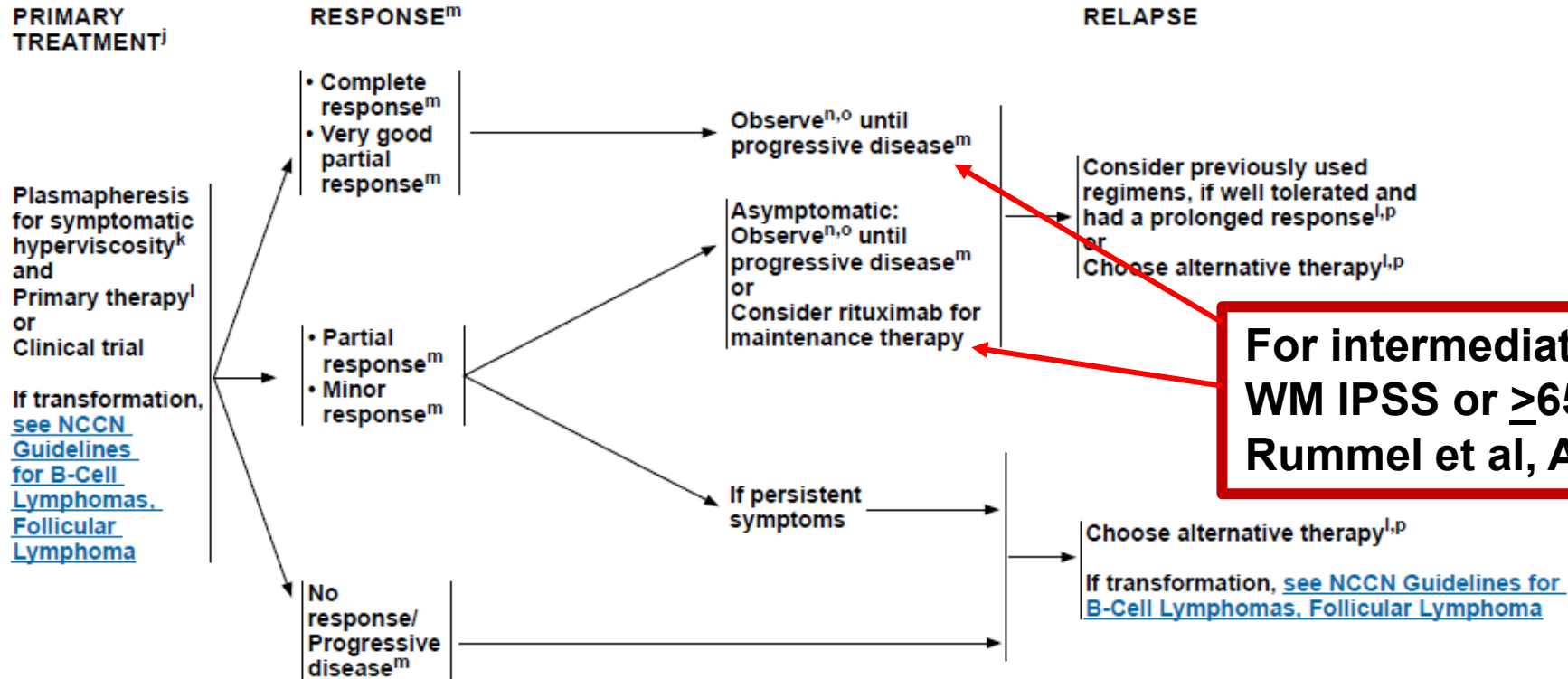
Most Frequent AEs, n (%)	Grade 1-2	Grade 3	Grade 4
Headache	41 (39)	0	0
Diarrhea	33 (31)	2 (2)	0
Contusion	31 (29)	0	0
Dizziness	27 (25)	0	0
URTI	23 (22)	0	0
Fatigue	22 (21)	2 (2)	0
Nausea	22 (21)	2 (2)	0
Constipation	22 (21)	0	0
Arthralgia	20 (19)	1 (1)	0
Back pain	18 (17)	1 (1)	0
Cough	18 (17)	0	0
Pyrexia	17 (16)	1 (1)	0
Vomiting	17 (16)	1 (1)	0
Rash	16 (15)	0	0

- Atrial fibrillation occurred in 5% (5/106) of patients
 - All events were grade 1-2 except for one (1%) grade 3 event
- Hypertension occurred in 5% (5/106) of patients
- 28% (30/106) of patients discontinued acalabrutinib during the study period
 - AEs led to discontinuation in 7% (7/106) of patients



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Waldenström Macroglobulinemia/ Lymphoplasmacytic Lymphoma



For intermediate/high WM IPSS or ≥65 years old Rummel et al, ASH 2019

^j Intent of therapy should be based on palliation of symptoms, not necessarily levels of IgM unless the patient is exhibiting evidence of symptomatic hyperviscosity.

^k Plasmapheresis should be performed for patients with symptomatic hyperviscosity, and before treatment with rituximab-containing regimen in patients with IgM ≥4000 mg/dL. IgM should be monitored closely in these patients thereafter and plasmapheresis should be considered again if symptomatic hyperviscosity recurs or if IgM is ≥4000 mg/dL while on rituximab-containing therapy. RBC transfusion, if indicated, should be done after plasmapheresis to prevent added hyperviscosity load.

^l See [Waldenström Macroglobulinemia/Lymphoplasmacytic Lymphoma Therapy \(WM/LPL-B\)](#).

^m See [Response Criteria for WM/LPL \(WM/LPL-C\)](#).

ⁿ See [NCCN Guidelines for Survivorship](#).

^o CBC, complete metabolic panel, and IgM every 3 months for 2 years, then every 4–6 months for additional 3 years, then every 6–12 months. Progression based on IgM levels alone, without symptoms, should not be reason to retreat.

^p Caution should be used when re-treating with myelosuppressive regimens due to cumulative toxicities.

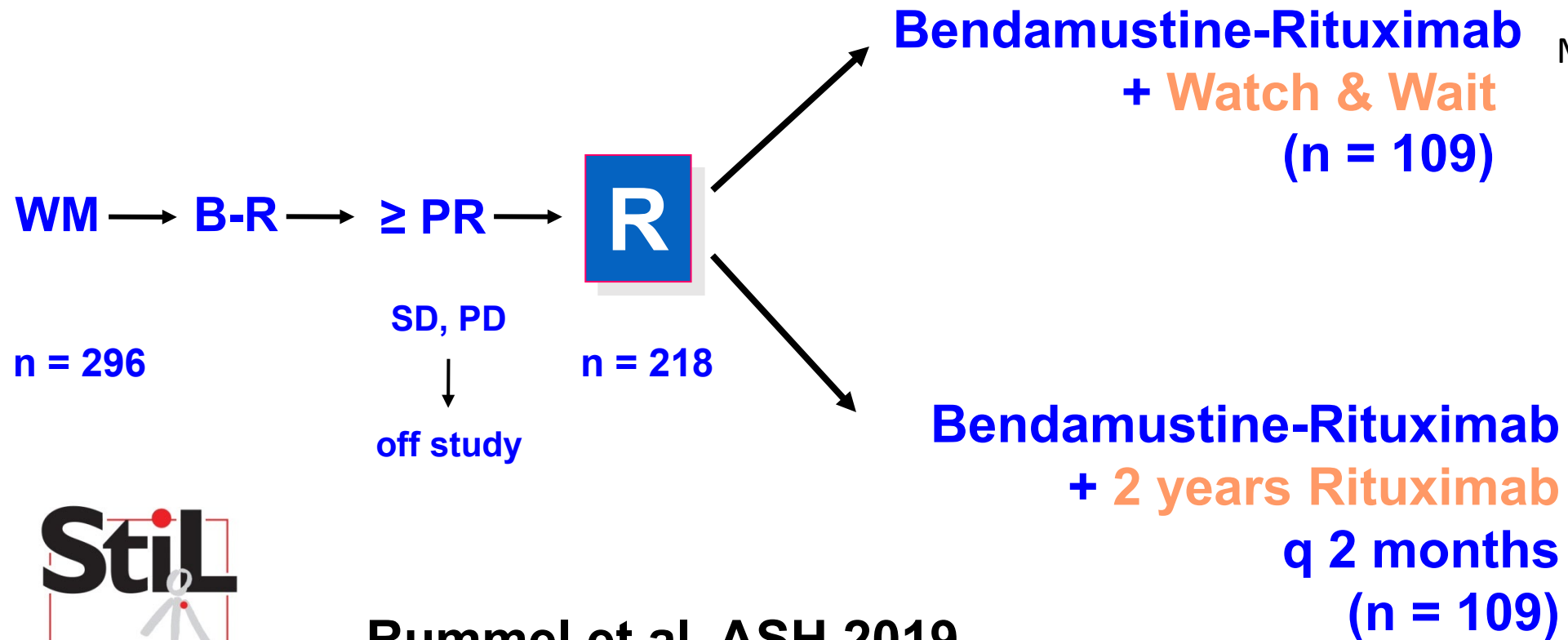
Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

B-R + Watch & Wait vs. B-R + 2 years Rituximab



M. Rummel

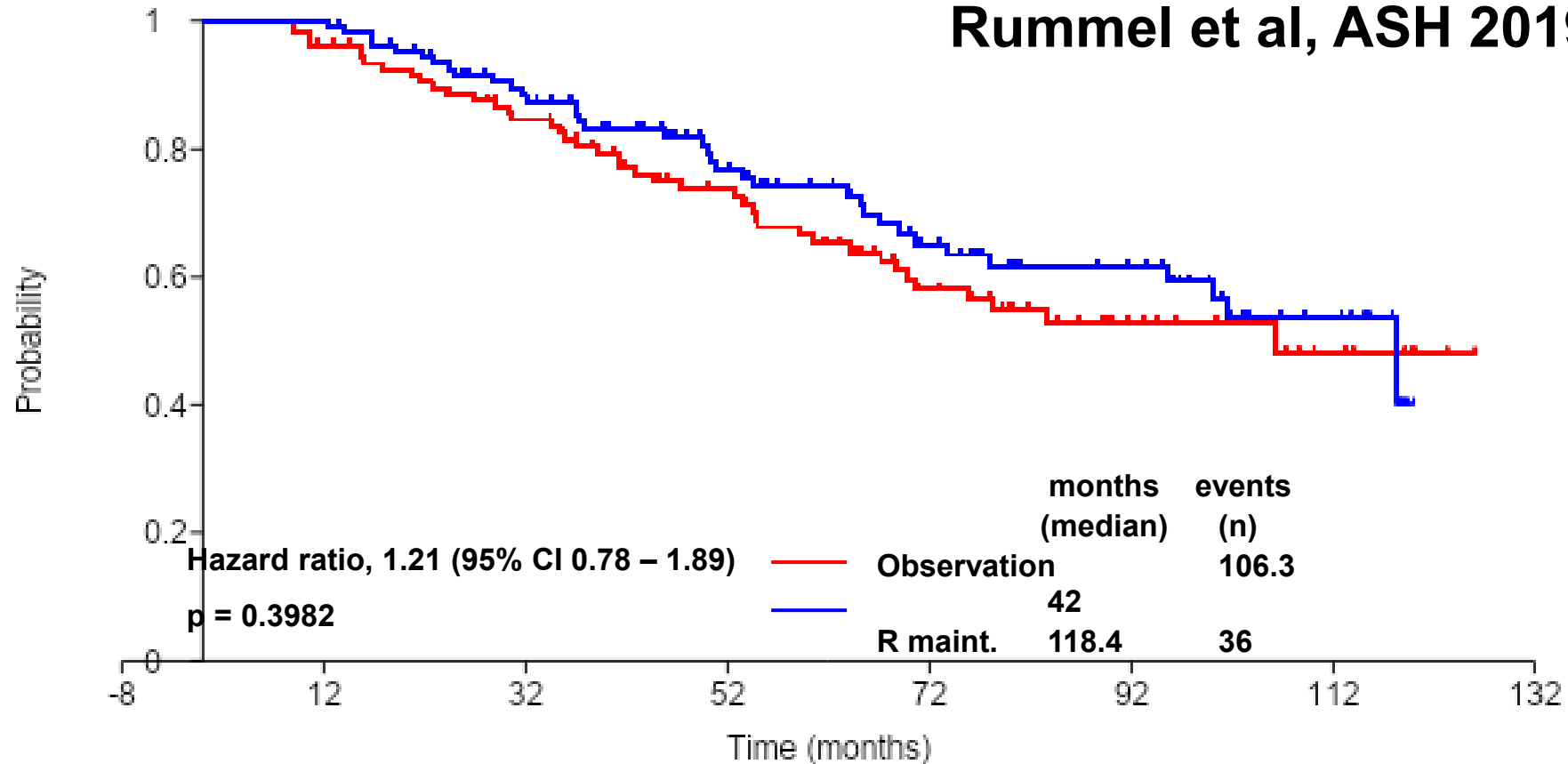
StiL NHL 7-2008 - MAINTAIN



Rummel et al, ASH 2019

Progression free survival (80 months median follow-up)

Rummel et al, ASH 2019

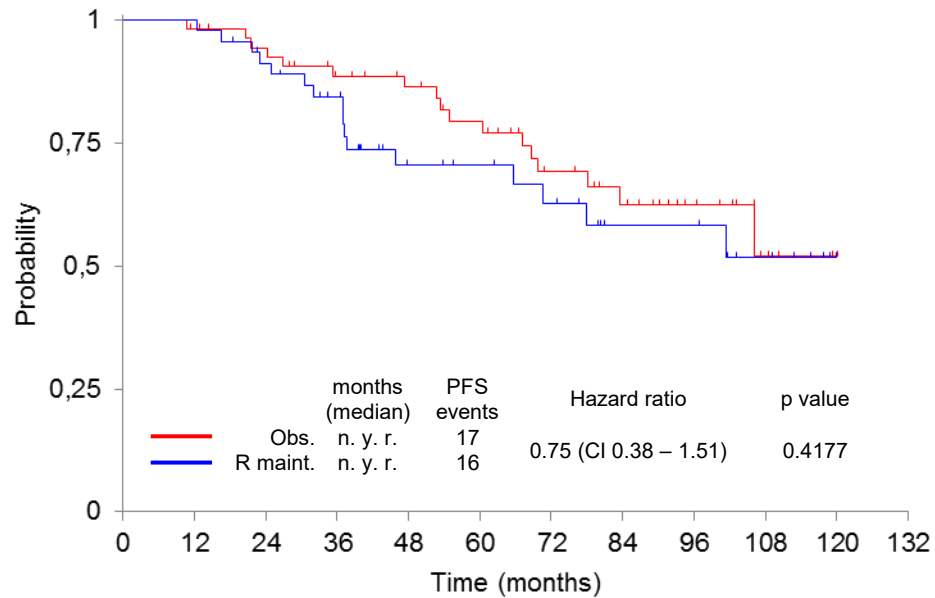


Pts at risk		12	32	52	72	92	112	132	
Observ.	109	102	92	79	62	54	39	27	18
R maint.	109	109	96	83	65	52	41	30	25

PFS: Patient age

Rummel et al, ASH 2019

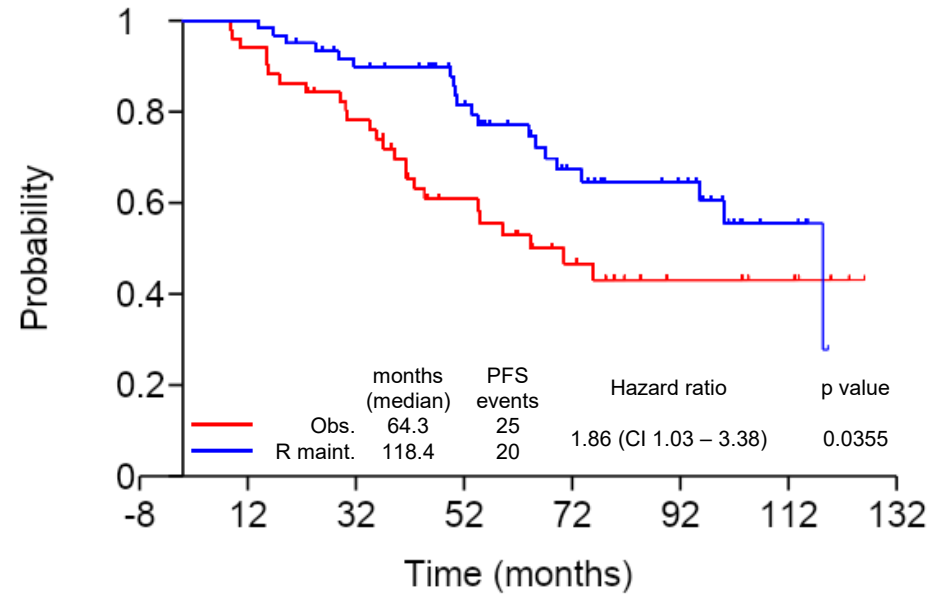
Patients younger than 65 years



Hazard ratio 0.75 (95% CI 0.38 – 1.51)

p = 0.4177

Patients older than 65 years



Hazard ratio 1.86 (95% CI 1.03 – 3.38)

p = 0.0355

IWWM-2 Workshop Classification of WM, IGM MGUS, and IgM Related Disorders.

	IgM Monoclonal Protein ¹	Histological Infiltration by LPL ²	Symptomatic ³
IgM MGUS	+	-	-
IgM Related Disorders	+	-	+
Asymptomatic WM	+	+	-
Symptomatic WM	+	+	+

1. IgM monoclonal gammopathy of any concentration
2. Bone infiltration by small lymphocytes, plasmacytoid cells and plasma cells. Any level of detectable infiltrate by histological examination. Flow or molecular disease detection does not fulfill WM diagnostic criteria.
3. Symptomatic Status defined by IWWM-2 consensus criteria and indicative of need for treatment. Kyle et al, Semin Oncol. 2003.

Morbidities mediated by monoclonal IgM and associated light chains in WM.

Property of IgM monoclonal protein	Diagnostic condition	Clinical manifestations
Pentameric structure	Hyperviscosity	Headaches, blurred vision, epistaxis, retinal hemorrhages, leg cramps, impaired mentation, intracranial hemorrhage
Precipitation on cooling	Cryoglobulinemia (type I)	Raynaud phenomenon, acrocyanosis, ulcers, purpura, cold urticaria
Autoantibody activity to myelin-associated glycoprotein (MAG), ganglioside M1 (GM1), sulfatide moieties on peripheral nerve sheaths	Peripheral neuropathies	Sensorimotor neuropathies, painful neuropathies, ataxic gait, bilateral foot drop
Autoantibody activity to IgG	Cryoglobulinemia (type II)	Purpura, arthralgias, renal failure, sensorimotor neuropathies
Autoantibody activity to red blood cell antigens	Cold agglutinins	Hemolytic anemia, Raynaud phenomenon, acrocyanosis, livedo reticularis
Tissue deposition as amorphous aggregates	Organ dysfunction	Skin: bullous skin disease, papules, Schnitzler syndrome; GI: diarrhea, malabsorption, bleeding; kidney: proteinuria, renal failure (light chain component)
Tissue deposition as amyloid fibrils (light chain component most commonly)	Organ dysfunction	Fatigue, weight loss, edema, hepatomegaly, macroglossia, organ dysfunction of involved organs: heart, kidney, liver, and peripheral sensory and autonomic nerves

IGM MGCS treated under WM/LPL NCCN guidelines

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NCCN Guidelines Version 1.2022
Multiple Myeloma

[NCCN Guidelines Index](#)
[Table of Contents](#)
[Discussion](#)

MONOCLONAL GAMMOPATHY OF CLINICAL SIGNIFICANCE (FOR MGRS, SEE MGRS-1)

MONOCLONAL GAMMOPATHY OF NEUROLOGICAL SIGNIFICANCE

INITIAL WORKUP	CLINICAL FINDINGS
<p>IgM^κ MGNS (Monoclonal gammopathy of neurological significance) suspected</p> <ul style="list-style-type: none"> • Rule out other causes of neuropathy <ul style="list-style-type: none"> ▶ Diabetes ▶ Cobalamin deficiency ▶ Thyroid dysfunction ▶ Lyme disease ▶ HIV infection ▶ Syphilis ▶ Autoimmune disease ▶ Cryoglobulinemia • Evaluation for light chain amyloidosis, (See NCCN Guidelines for Systemic Light Chain Amyloidosis), WM (See NCCN Guidelines for WM/LPL), or POEMS (See POEMS-1), if appropriate. • Anti-MAG antibodies^a • Ganglioside antibody panel • Nerve conduction study (NCS)/electromyogram (EMG)^b • Neurology consult • MYD88,^b L265P allele-specific PCR (AS-PCR) testing of bone marrow • Chest/abdominal/pelvic CT with contrast when possible <p>Useful in certain circumstances</p> <ul style="list-style-type: none"> • Sural nerve biopsy • CXCR4 gene mutation testing 	<p>High suspicion</p> <ul style="list-style-type: none"> • Sensory predominant • Length dependent • Slow progression (years) • Bilateral and symmetrical • Antibodies present • Demyelination by EMG/NCS • OR intermediate suspicion (not high or low suspicion) AND affecting activities of daily living (ADLs) <p>See NCCN Guidelines for Waldenström Macroglobulinemia/Lymphoplasmacytic Lymphoma</p> <p>→ Observation</p> <p>Low suspicion</p> <ul style="list-style-type: none"> • Motor/pain predominant • Non-length dependent • Rapid progression (weeks to months) • Unilateral/asymmetrical • Antibodies not present • No demyelination by EMG/NCS • OR intermediate/high suspicion AND not affecting ADLs

^a In patients presenting with suspected disease related to peripheral neuropathy, rule out amyloidosis in patients presenting with nephrotic syndrome or unexplained cardiac problems.
^b MYD88 wild-type occurs in <10% of patients and should not be used to exclude diagnosis of WM if other criteria are met.

Note: All recommendations are category 2A unless otherwise indicated.
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MGRS-1

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NCCN Guidelines Version 1.2022
Multiple Myeloma

[NCCN Guidelines Index](#)
[Table of Contents](#)
[Discussion](#)

MONOCLONAL GAMMOPATHY OF CLINICAL SIGNIFICANCE

MONOCLONAL GAMMOPATHY OF RENAL SIGNIFICANCE

CLINICAL FINDINGS	INITIAL WORKUP	ADDITIONAL WORKUP
<p>MGRS (Monoclonal gammopathy of renal significance) suspected</p>	<p>Evaluate for kidney disease</p> <ul style="list-style-type: none"> • Kidney function: eGFR • Urinalysis • Metabolic testing 	<p>Renal biopsy recommended if:</p> <ul style="list-style-type: none"> • AKI stage 3 • eGFR <60 mL/min and >2mL/min per year decline • Proteinuria (>1 g/day) • Albumin:creatinine >30 mg/mmol • Fanconi syndrome <p>Consider renal biopsy if:</p> <ul style="list-style-type: none"> • AKI stage 1 or 2 • eGFR <60 mL/min and <2 mL/min per year decline • Albumin:creatinine 3–30 mg/mmol and GFR <60 mL/min • Evidence of light chain proteinuria <p>Defer renal biopsy if:</p> <ul style="list-style-type: none"> • Stable eGFR • Normal urinalysis • No evidence of light chain proteinuria

To confirm diagnosis of MGRS:

- Light microscopy
- Immunofluorescence staining for IgG subclasses, IgA and IgM, and kappa and lambda

Note: M protein detected in serum and/or urine must match the one found in the renal biopsy

- Electron microscopy
- PET/CT, low-dose CT, or whole-body MRI as clinically indicated
- Bone marrow biopsy if suspected to have MM or WM

Additional workup as clinically indicated:

- FISH panel for myeloma and polymerase chain reaction (PCR) assay for MYD88 L265P
- Excisional lymph node biopsy, if other B-cell lymphomas are suspected
- Peripheral blood flow cytometry for diagnosis of CLL (See [NCCN Guidelines for Chronic Lymphocytic Leukemia/Small Lymphocytic Lymphoma](#))
- Evaluate for light chain amyloidosis (See [NCCN Guidelines for Systemic Light Chain Amyloidosis](#))

→ For management See [MGRS-2](#)

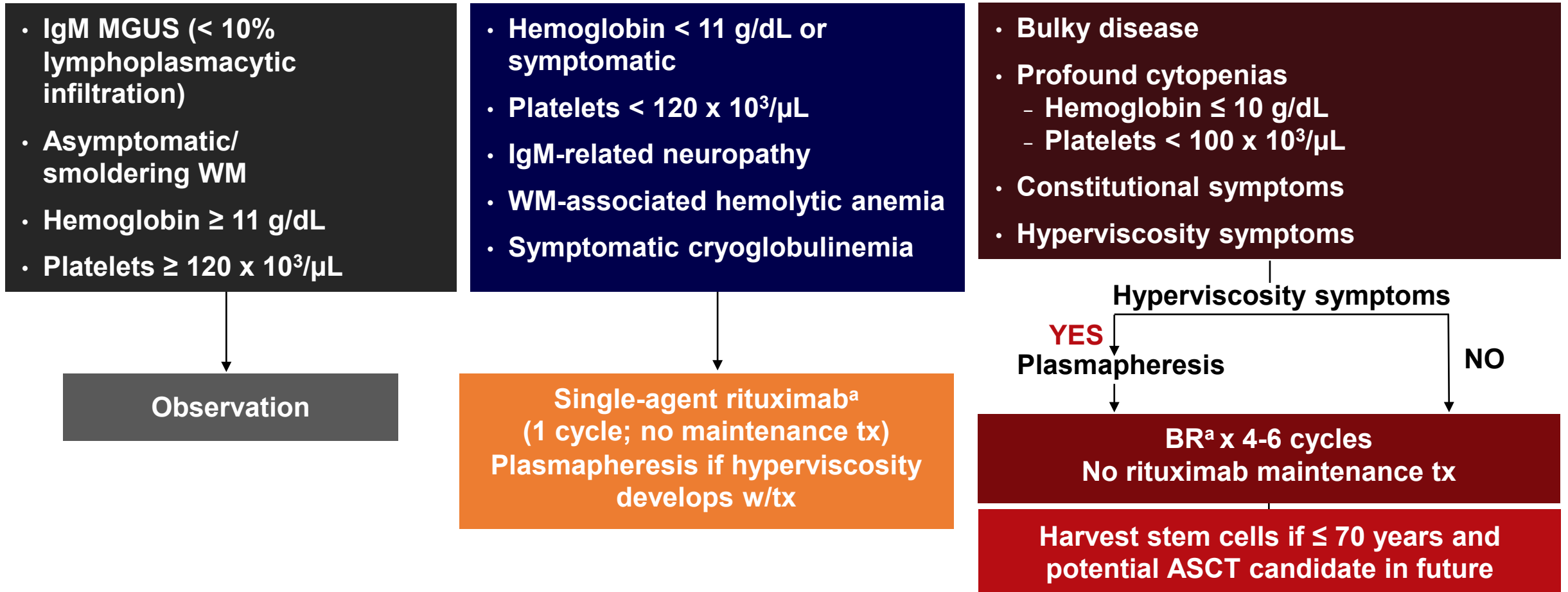
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MGRS-1

Mayo Stratification of WM and Risk-Adapted Therapy Guidelines

Consensus for Newly Diagnosed WM



^aSix cycles of DRC is an alternative if the disease burden is low.

Mayo Stratification of WM and Risk-Adapted Therapy Guidelines

WM Consensus for Off-Duty Salvage Therapy

Time to next therapy \geq 3 years from previous therapy

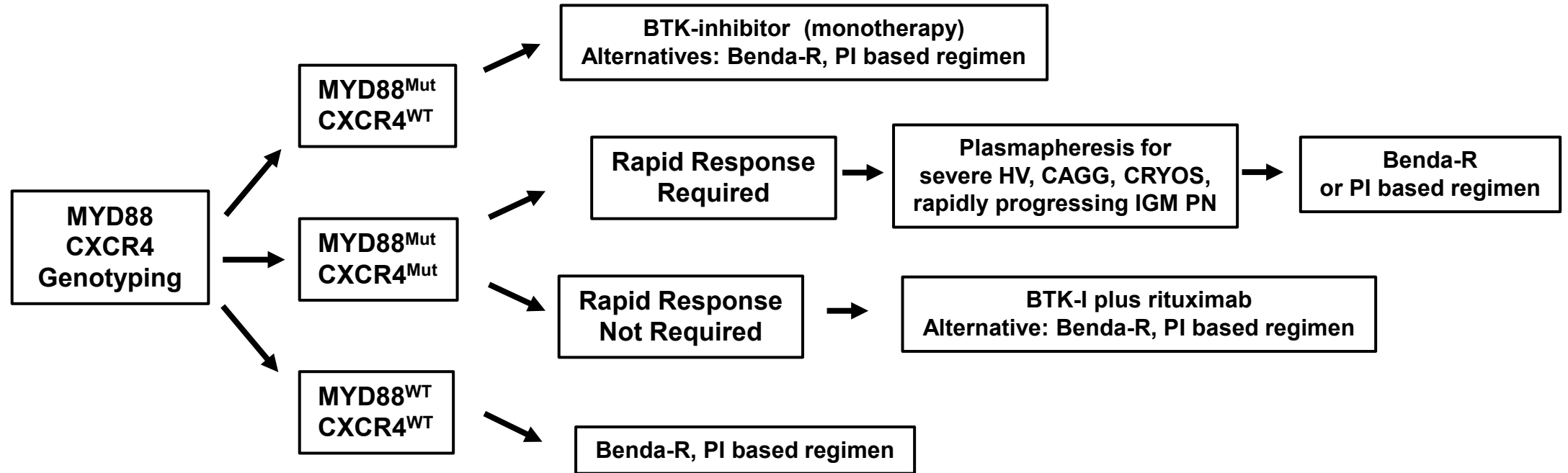
Repeat original therapy

- Ibrutinib monotherapy^a
- BDR if preexisting peripheral neuropathy Grade < 2
- DRC
- BR

ASCT in select patients

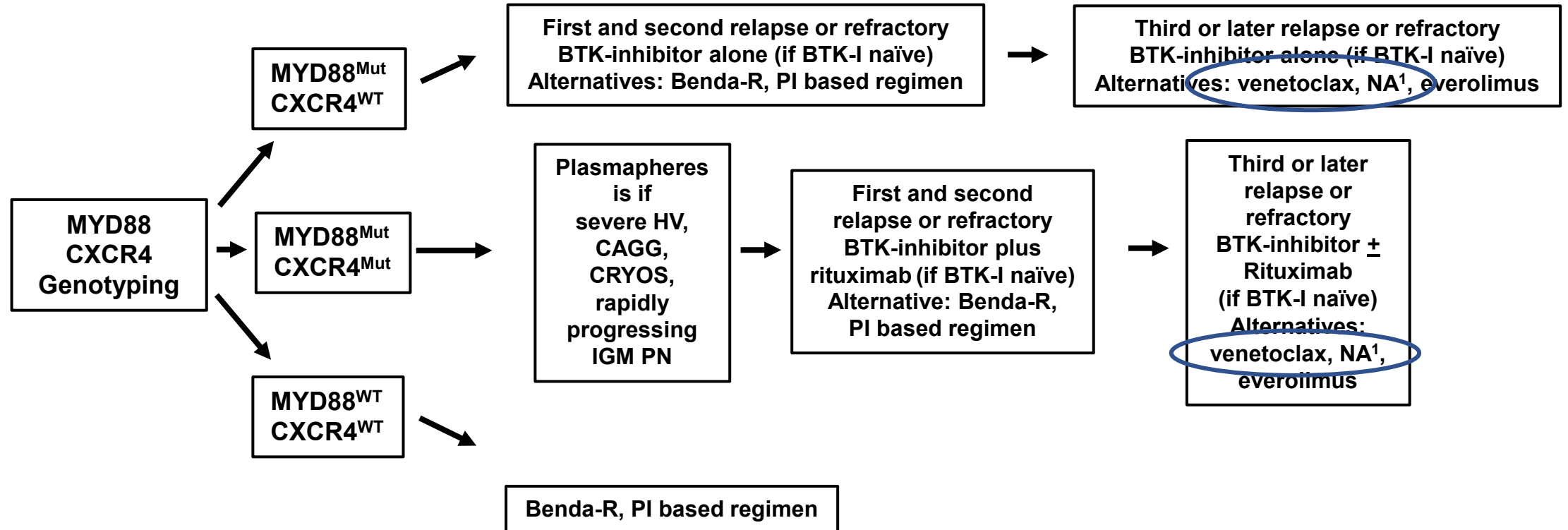
^aIf not previously used.

Genomic Based Treatment Approach to Symptomatic Treatment Naïve WM



- Rituximab should be held for serum IgM $\geq 4,000$ mg/dL
- Benda-R for bulky adenopathy or extramedullary disease.
- PI based regimen for symptomatic amyloidosis, and possible ASCT as consolidation.
- Rituximab alone, or with ibrutinib if MYD88^{Mut} or bendamustine for IgM PN depending on severity and pace of progression.
- Maintenance rituximab may be considered in patients responding to rituximab based regimens.

Genomic Based Treatment Approach to Symptomatic Relapsed or Refractory WM



- Nucleoside analogues (NA) should be avoided in younger patients, and candidates for ASCT.¹
- ASCT may be considered in patients with multiple relapses, and chemosensitive disease.

11th International Workshop for Waldenstrom's Macroglobulinemia Madrid, October 6-8, 2022



www.waldenstromsworkshop.org

